7. CENOZOIC DIATOM BIOSTRATIGRAPHY AND PALEOCEANOGRAPHY OF THE **ROCKALL PLATEAU REGION, NORTH ATLANTIC, DEEP SEA DRILLING PROJECT LEG 81**¹

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ABSTRACT

Diatoms are present within the Eocene to Holocene sediments recovered during Deep Sea Drilling Project (DSDP) Leg 81. A late Cenozoic, North Atlantic diatom zonation is developed as a result of the occurrence of common wellpreserved diatoms within middle Miocene to Holocene sediments. This high-latitude diatom zonation correlates with the zonation defined for the low-latitude Pacific.

Two distinct intervals of dissolution are recognized in Leg 81 sediments, a late-Miocene interval associated with the first occurrence of Thalassiosira convexa and T. miocena (6.2 m.y. ago) and a Pliocene-Pleistocene interval associated with oceanographic responses to the onset of Northern Hemisphere glaciation. Detailed analyses of the uppermost 50 m of Hole 552A indicate that diatom abundance within sediments of the Rockall Region is primarily controlled by fluctuations in productivity resulting from latitudinal migration of water masses.

INTRODUCTION

Biostratigraphic and paleoecological studies of diatoms recovered from deep-sea sediments within the North Atlantic are quite limited when compared to the quantity of literature available concerning the diatom flora present in sediments from the North Pacific.

Few siliceous microfossil studies have been completed in the polar region of the North Atlantic. Gran (1904) published a taxonomic study of the phytoplankton of the Arctic Sea. More recently, the biostratigraphic and taxonomic studies of Schrader and Fenner (1976) and Dzinoridze et al. (1976) have been completed from sediments recovered during DSDP Leg 38 within the Norwegian-Greenland Seas.

Studies of the diatom flora in the northwestern Atlantic have been completed by Bukry (1980) for Eocene material recovered from DSDP Sites 417 and 418, Schrader (1979) for Miocene material recovered from DSDP Sites 407 and 408, and Bukry (1978a) for sediments recovered from DSDP Leg 44.

Diatoms in the surface waters of the eastern North Atlantic and the English Channel Region were recorded by Cleve (1900). Other studies within the mid-latitude Atlantic include Lohman (1941), Maynard (1976), and Pokras (1981), in which surface sediments throughout the high-latitude Atlantic were examined for fluctuations in the Pleistocene diatom assemblage. The middle Miocene diatom flora from sediments along the Atlantic margin of North America were documented by Abbott (1978) and Andrews (1978).

Numerous workers have examined the diatom flora from the equatorial Atlantic. These include the studies of Riely (1957), Kolbe (1957), Chyong Ngok (1971), Schrader (1977), and Fenner (1977). In addition, Hass and Schrader (1979) and Schuette and Schrader (1981) have examined the phytoplankton within the upwelling region off northwest Africa. Furthermore, Wornardt (1974), Burckle (1977), Schrader and Gersonde (1978), Gersonde (1979, 1980), and Gardette (1979) have studied the diatom biostratigraphy and paleoecology of the Messinian Stage within the Mediterranean Region.

During DSDP Leg 81, four sites were drilled within the western margin of the Hatton Rockall Plateau (see Fig. 1). The presence of common diatoms in the middle to upper Miocene sediments of Site 555 and the upper Miocene to Holocene sediments of Hole 552A allows correlation of these sites with lower latitudes. Furthermore, the recovery of a well-preserved Pliocene-Pleistocene interval containing diatoms at Hole 552A makes possible the reconstruction of the paleoceanography of the North Atlantic for approximately the last 2.8 m.y.

ZONATION

Diatom zonations previously established within the mid- and high-latitude North Atlantic consist of zonal schemes developed by Schrader and Fenner (1976) and Dzinoridze et al. (1976) for the Norwegian-Greenland Sea Region, and Abbott (1978) for the continental shelf region of eastern North America. Correlation of these zones appears in Figure 2.

Although the Eocene to Holocene zonation developed by Schrader and Fenner (1976) is partially based on midlatitude oceanic species, the majority of the species used as zonal indicators are endemic to the Arctic Region. These species are not present within the Rockall Plateau Region and therefore only allow partial recognition of Schrader and Fenner's zonation.

Dzinoridze et al. (1976) developed a zonation defined by high-latitude forms such as Denticulopsis hustedtii, D. lauta, D. kamtschatica, D. seminae, and Thalassiosira zabelinae. Of these species, only D. hustedtii and D. lauta consistently occur within the warm-water diatom assemblage that is present in the Rockall Region, there-

¹ Roberts, D. G., Schnitker, D., et al., Init. Repts. DSDP, 81: Washington (U.S. Govt. Printing Office). ² Address: U.S. Geological Survey, 345 Middlefield Road, Menlo Park, CA 94025.



Figure 1. Location of DSDP sites cored during Leg 81.

fore also rendering their zonation only partially useful in the warmer waters of the Rockall Region.

Similarly, the zonation developed by Abbott (1978) for the Miocene strata along the Atlantic margin of North America is only partially useful in the Rockall Region. This zonation is based primarily on shallow-water coastal forms that do not occur within Leg-81 material.

Diatoms recovered during Leg 37 (Schrader, 1977) in the low-latitude North Atlantic comprise a tropical assemblage including the species *Rhizosolenia praebergonii*, *Nitzschia miocenica*, and *N. porteri*. The diatom biostratigraphy of Leg-37 sediments is based on the diatom zonations developed by Burckle (1969, 1972).

The warm temperate diatom assemblage present within the late Neogene sediments recovered during Leg 81 also allows recognition and partial usage of the Partial Range Diatom Zones defined by Burckle (1969, 1972) for upper Miocene to Holocene sediments of the equatorial Pacific. Although the majority of the index species used to define Burckle's zones are present in the sediments recovered from the Rockall Plateau Region, the extremely rare occurrence of *R. praebergonii* and *N. miocenica* and the preservational or ecological exclusion of *N. porteri* require that Burckle's zonal scheme be modified to be useful within the Hatton-Rockall Region of the North Atlantic (see Fig. 3).

The middle Miocene interval at Site 555 contains wellpreserved diatoms and thus allows the development of a middle Miocene diatom zonation. The development of this zonation is based on the ranges of selected species present at both Sites 555 and 408 (Reykjanes Ridge, see Figs. 1 and 4).

Coscinodiscus lewisianus Interval

Author: Base, not defined; Top, Schrader (1976).

Definition: The base of this interval is not defined. The top of this interval is placed at the last occurrence of *C. lewisianus*.

Remarks. The Coscinodiscus lewisianus Interval is recognized at DSDP Sites 555, 408, and 338. No base is presently defined for this interval because of dissolution at both Sites 555 and 408. However, lower middle Miocene sediments at Site 338 contain well-preserved diatoms that stratigraphically range from the first occurrence of *Denticulopsis lauta* to the last occurrence of *C. lewisianus.* Schrader and Fenner (1976) assigned Cores 338-8 through 338-11 to their middle Miocene Norwegian Sea Zones (*Actinocyclus ingens* to *Rhizosolenia bulbosa* Zones) which are based primarily on species endemic to the Norwegian Sea Region.

Recently Barron (1980a, 1981) developed a zonation that has greater geographic applicability and can be recognized at Site 338. The presence of D. lauta, D. hyalina, D. hustedtii, A. ingens, Raphidodiscus marylandicus, C. plicatus, and C. lewisianus allows Cores 338-8 through 338-11 to be assigned to the Denticulopsis lauta and the Denticulopsis hustedtii-Denticulopsis lauta Zones of Barron (1980a, 1981). The Denticulopsis lauta Zone extends from the first occurrence of D. lauta in Sample 338-11-4, 85-86 cm to the first occurrence of D. hustedtii recorded by Schrader and Fenner (1976) in Sample 338-11-2, 85-86 cm. The first occurrence of D. hyalina, which defines the Denticulopsis lauta Subzone "a"/Subzone "b" boundary, is recorded in Sample 338-11-3, 85-86 cm, although the first occurrence of D. lauta is recorded in Sample 338-11-4, 85-86 cm. The first occurrence of D. punctata, A. ingens, and Mederia splendida is also in this sample, suggesting that an unconformity occurs directly below this sample and that the true first occurrence of D. lauta is missing.

Barron (1980a, 1981) defined four subzones for the *Denticulopsis hustedtii-Denticulopsis lauta* Zone. However, only Subzone "a" defined as the interval from the first occurrence of *D. hustedtii* to the first occurrence of *D. praedimorpha* is recognized at Site 338. Within the uppermost portion of this subzone, the first occurrence of *C. plicatus* and the last occurrence of *C. lewisianus* are observed (Barron, 1981).

The first occurrence of D. hustedtii is recorded in Sample 338-11-2, 85-86 cm. The first occurrence of C. plicatus and the last occurrence of C. lewisianus both occur in Sample 338-8-2, 10-11 cm. Because diatoms are barren above this sample the true last occurrence of C. lewisianus is uncertain. However, the first occurrence of C. plicatus within this sample suggests that the true last occurrence of C. lewisianus should occur only slightly above this sample.

Other useful markers within Subzone "a" are the last occurrence of R. marylandicus and the transition from common D. hyalina to common D. hustedtii. Both are recognized at Site 338. The last R. marylandicus is re-

CENOZOIC DIATOMS BIOSTRATIGRAPHY AND PALEOCEANOGRAPHY

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Figure 2. Correlation of the Leg 81 diatom zonation modified from Burckle (1972) with the high-latitude Norwegian Sea diatom zonations of Schrader and Fenner (1976) and Dzinoridze et al. (1976), the middle Miocene western Atlantic margin diatom zonation of Abbott (1978), and the low-latitude Pacific diatom zonation of Burckle (1972).



Figure 3. Correlation of the Leg 81 diatom zonation and datum levels with the equatorial Pacific diatom zonation defined by Burckle (1972).

corded in Sample 338-11-1, 65-66 cm, and the *D. hy-alina-D. hustedtii* transition occurs at approximately Sample 338-10-1, 135-136 cm.

The ability to distinguish these zones in the Norwegian Sea allows the base of the *Coscinodiscus lewisianus* Interval to be tentatively placed at the first occurrence of *D. hustedtii*. However, further correlations are required to determine if this datum is time equivalent between high and middle latitudes of the Atlantic. This is especially critical since the first occurrence of *D. lauta* is diachronus between these latitudes.

D. lauta first occurs stratigraphically above the last occurrence of C. lewisianus (see Fig. 4) at both Sites 555 and 408. This stratigraphic position is similar to the first occurrence of D. lauta at mid-latitudes. The first occurrence of D. lauta at Site 338 is recorded stratigraphically below the first C. lewisianus. This stratigraphic succession is similar to that of the high-latitude Pacific.

Other floral elements: A. ingens, C. obscurus, C. plicatus, C. tabularis, C. vetustissimus, Craspedodiscus coscinodiscus, D. hustedtii, D. nicobarica, D. punctata, Rhizosolenia miocenica, R. praebarboi.

Correlation: The top of this interval approximates the middle Miocene NN6/NN7 calcareous nannofossil zonal boundary as used by Backman (this volume). The *Coscinodiscus lewisianus* Interval also correlates to Subzone "a" of the *Denticulopsis hustedtii-Denticulopsis lauta* Zone of Barron (1980a) to North Pacific Diatom Zone XXI (NPDZ XXI) of Schrader (1973), and to the *Coscinodiscus lewisianus* Zone of Barron (in press).

Denticulopsis nicobarica Zone

Author: Base, Schrader (1976); Top, Baldauf (this chapter).

Definition: The base of this zone is defined by the last occurrence of *Coscinodiscus lewisianus*. The top is defined by the first occurrence of *Rhizosolenia barboi*.

Remarks: This zone is recognized at both Sites 555 and 408. Useful datums within this zone include the first occurrence of *Denticulopsis praedimorpha* and the last occurrence of *D. nicobarica*. Schrader (1976) defined the top of his *Denticulopsis nicobarica* Zone by the first occurrence of *D. hustedtii*. This top has been modified here.

Other floral elements: Actinocyclus ingens, Asterolampra marylandica, C. plicatus, C. radiatus, Rossiella paleacea, D. praedimorpha, D. hustedtii, D. lauta, D.



Figure 4. Comparison of stratigraphically useful datums between DSDP Sites 408 and 555.

punctata, D. punctata var. hustedtii, Rhizosolenia praebarboi, Rouxia californica, and Thalassiothrix miocenica.

Correlation: The top of this zone correlates with the lowest portion of the calcareous nannofossil Zone NN9 as recognized by Backman (this volume). The *Denticulopsis nicobarica* Zone correlates to Subzone "a" and "b" of the *Denticulopsis hustedtii–Denticulopsis lauta* Zone of Barron (1980a), NPDZ XIX and XX of Schra-

der (1973), and the Coscinodiscus gigas var. diorama Zone of Barron (in press).

Rhizosolenia barboi Zone

Author: Baldauf (this chapter).

Definition: The base of this zone is defined by the first occurrence of *R*. *barboi*. The top of this zone is

placed at the last occurrence of *Denticulopsis punctata* var. *hustedtii*.

Remarks: This zone is recognized at both Sites 555 and 408. The first occurrence of *Hemidiscus cuneiformis* at Site 555 is similar to its first occurrence within the mid-latitude Pacific (Barron, 1981) and is a useful datum within this zone. The top of this zone approximates the middle Miocene/late Miocene boundary.

Other floral elements: Actinocyclus ingens, Coscinodiscus salisburyanus, C. yabei, Craspedodiscus coscinodiscus, Coscinodiscus plicatus, D. hustedtii, D. lauta, D. praedimorpha, and R. praebarboi.

Correlation: This zone correlates to the lowest portion of the *Coscinodiscus yabei* Zone of Burckle (1972), Subzone "c" of the *Denticulopsis hustedtii–Denticulopsis lauta* Zone of Barron (1980a), and the *Craspedodiscus coscinodiscus* Zone of Barron (in press).

Denticulopsis praedimorpha Zone

Author: Baldauf (this chapter).

Definition: The base of this zone is defined by the last occurrence of *D. punctata* var. *hustedtii*. The top is defined by last occurrence of *D. praedimorpha*.

Remarks: The last occurrence of D. punctata occurs within this zone at both Sites 555 and 408. However, the range of D. punctata is stratigraphically higher at Site 555, where the last occurrence approximates the last occurrence of D. praedimorpha. The last occurrence of D. punctata at Site 408 occurs in the middle portion of the Denticulopsis praedimorpha Zone. Therefore, the reliability of D. punctata as a secondary datum is doubtful. Useful datums within this zone include the first occurrence of Thalassiosira eccentrica and the last occurrence of Actinocyclus ingens, both of which occur within the upper portion of this zone. The last occurrence of A. ingens within this zone is equivalent to the range of this species reported from temperate latitudes of the Pacific (Schrader, 1973, and Barron, 1980a) rather than to the range of this species reported from tropical regions (see Barron, 1981).

Other floral elements: Coscinodiscus plicatus, C. salisburyanus, C. tabularis, C. vetustisimus, C. yabei, D. hyalina, Hemidiscus cuneiformis, Nitzschia praereinholdii, Rhizosolenia barboi, and Synedra jouseana.

Correlation: This zone correlates to the lower portion of the *Coscinodiscus yabei* Zone of Burckle (1972) and to Subzone "d" of the *Denticulopsis hustedtii–Denticulopsis lauta* Zone of Barron (1980a).

Coscinodiscus yabei Zone

Author: Base, Baldauf (this chapter); Top, Burckle (1972).

Definition: The base of this zone is defined by the last occurrence of *Denticulopsis praedimorpha*. The top of this zone is placed at the last occurrence of *C. yabei*.

Remarks: Although the top of this zone is defined by the last occurrence of C. *yabei*, the range of this species is diachronous between subtropical and temperate assemblages (see Barron, 1980a). At Site 555 the range of C. *yabei* is comparable to a temperate range; that is the

range is similar to the range of *C. yabei* at North Pacific (Barron, 1980a) Site 173. The last occurrence of *C. yabei* at these sites approximates the last occurrence of *D. praedimorpha* and is older than the last occurrence of *C. yabei* reported from the tropics by Burckle (1972). Within the tropical region, the last occurrence of *C. yabei* approximates the first occurrences of *Nitzschia fossilis, N. cylindrica* and *N. reinholdii* (Barron, written comm.). Schrader (1977) and Barron (1980a) report that the last occurrence of *C. yabei* within tropical regions.

At Site 555 the last occurrence of *C. plicatus* supports this stratigraphic range since its last occurrence approximates the first occurrence of the younger *Nitzschia* group. Therefore, in this study the last occurrence of *C. plicatus* is used as the primary datum in defining the top of the *Coscinodiscus yabei* Zone. The top of this zone is not represented at Site 408 as a result of dissolution. At Sites 552, 553, and 554, dissolution also inhibits the recognition of this zone. However, at Site 554 several cores contain sparse diatoms that are tentatively placed within the *Coscinodiscus yabei* Zone based on the co-occurrence of the primitive *Nitzschia* group (*N. praefossilis–N. praereinholdii*) and *Thalassiosira eccentrica* without the *Denticulopsis* group. *C. plicatus* is presumed to be preservationally excluded.

Other floral elements: Actinocyclus ellipticus, C. radiatus, Hemidiscus cuneiformis, Melosira sulcata, and Rhizosolenia barboi.

Correlation: The top of this zone correlates with the lower portion of the *D. petterssonii* radiolarian zone as used by Westberg-Smith and Riedel (this volume). The *Coscinodiscus yabei* Zone overlaps with Subzone "a" of the *Denticulopsis hustedtii* Zone and the upper part of Subzone "d" of the *Denticulopsis hustedtii–Denticulopsis lauta* Zones of Barron (1980a).

Nitzschia porteri-Nitzschia miocenica Interval

Author: Burckle (1972, 1977).

Definition: The base of this interval is defined by the last occurrence of *Coscinodiscus yabei*. The top of this interval is defined by the first occurrence of *Thalassiosira convexa* and *T. miocenica*.

Remarks: Burckle (1972, 1977) defined the Nitzschia porteri and the Nitzschia miocenica Zones based on the last occurrence of C. yabei and the first occurrences of N. miocenica and T. convexa (see Fig. 3). Other useful datums defined within these zones include the last occurrence of N. porteri and the first occurrence of T. praeconvexa. Within the Rockall Region N. porteri is preservationally or ecologically excluded and N. miocenica is extremely rare. This prevents the recognition of the first occurrence of N. miocenica, which defines the boundary between the Nitzschia porteri and Nitzschia miocenica Zones and dictates that these zones be combined. The Nitzschia porteri-Nitzschia miocenica Interval is distinguished by the presence of N. fossilis, N. cylindrica, T. eccentrica, and T. nativa. The first occurrence of T. praeconvexa occurs within the upper part of this zone.

Other floral elements: C. tabularis, Hemidiscus cuneiformis, Melosira sulcata, N. reinholdii, and Rhizosolenia barboi.

Correlation: The top of this interval correlates with the top of the calcareous nannofossil Zone NN11 as used by Backman (this volume) and to the lowermost portion of the *Stichocorys peregrina* radiolarian Zone as used by Westberg-Smith and Riedel (this volume). The *Nitzschia porteri–Nitzschia miocenica* Interval correlates with Subzone "a" of the *Denticulopsis kamtschatica* and the upper portion of Subzone "b" of the *Denticulopsis hustedtii* Zone of Barron (1980a).

Thalassiosira convexa Zone

Author: Burckle (1972).

Definition: The base of this zone is defined by the first occurrence of *T. convexa*. The top of this zone is defined by the first occurrence of *Nitzschia jouseae*.

Remarks: Three subzones are recognized by Burckle (1972). The top of Subzone "A" (the oldest) is placed at the last occurrence of *T. praeconvexa*. The top of Subzone "B" is defined by the last occurrence of *T. miocenica*, and the top of Subzone "C" is equivalent to the top of the zone. Other useful datums within this zone include the last occurrence of *T. heats of T. and the first occurrence of T. oestrupii*. The first occurrence of *T. miocenica* is a secondary datum for the base of this zone. The base of this zone approximates the top of a dissolution interval that is present at all sites.

Other floral elements: Coscinodiscus nodulifer, C. tabularis, Hemidiscus cuneiformis, N. cylindrica, N. fossilis, N. reinholdii, and N. marina.

Correlation: The top of this zone tentatively correlates with the calcareous nannofossil Zones NN13/NN14 as used by Backman (this volume). The *Thalassiosira convexa* Zone correlates with Subzones "a" and "b" of the *Denticulopsis kamtschatica* Zone of Barron (1980a).

Nitzschia jouseae Zone

Author: Base, Burckle (1972); top, Baldauf (this chapter).

Definition: The base of this zone is defined by the first occurrence of N. *jouseae*. The top of this zone is defined by the last occurrence of N. *jouseae*.

Remarks: The top of the *Nitzschia jouseae* Zone as defined by Burckle (1972) is based on the first occurrence of *Rhizosolenia praebergonii*, which in the eastern Pacific is restricted to the equatorial region (see Burckle, 1972; 1978a). Within the western Pacific, Koizumi (1968) and Burckle (1972) record *R. praebergonii* in upper Tertiary sediments off Japan. Locally, within the Rockall Region, the occurrence of *R. praebergonii* is extremely rare and is not useful as a zonal marker. Therefore the top of the *Nitzschia jouseae* Zone (Burckle, 1972) is extended and here defined by the last occurrence of *N. jouseae*. The last occurrence of *N. cylindrica* occurs directly above the base of this zone.

Other floral elements: Coscinodiscus nodulifer, C. tabularis, Hemidiscus cuneiformis, Melosira sulcata, N. fossilis, N. marina, N. reinholdii, Synedra ulna, Thalassiosira oestrupii, and T. convexa. **Correlation:** This zone correlates to the *Nitzschia jou*seae Zone and Subzone "A" of the *Rhizosolenia prae*bergonii Zone of Burckle (1972). The *Nitzschia jouseae* Zone also in part correlates to NPDZ VIII and IX of Schrader (1973).

Nitzschia marina Zone

Author: Baldauf (this chapter).

Definition: The base of this zone is defined by the last occurrence of *N. jouseae*. The top of the zone is defined by the first occurrence of *Pseudoeunotia doliolus*.

Remarks: This zone is recognized by the occurrence of *N. marina*, *N. fossilis*, *N. reinholdii*, *Hemidiscus cuneiformis*, and *Thalassiosira oestrupii*. The last occurrence of *T. convexa* occurs within this zone.

Other floral elements: Coscinodiscus nodulifer, C. tabularis, Ethmodiscus rex, Melosira sulcata, Rhizosolenia barboi, R. hebatata, and Stephanopyxis turris.

Correlation: This zone correlates to Subzones "B" and "C" of the *Rhizosolenia praebergonii* Zone of Burckle (1972), and in part to NPDZ V and VIII of Schrader (1973).

Pseudoeunotia doliolus Zone

Author: Burckle (1972).

Definition: The base of this zone is defined by the first occurrence of *P. doliolus*. The top ranges to the present day.

Remarks: Burckle (1972) defined two subzones based on the last occurrence of *Rhizosolenia praerbergonii*. However, since *R. praebergonii* is so rare in Leg 81 material, no subzones are recognized. *R. curvirostris* occurs within the upper portion of this zone at both Hole 552A and Site 554.

Other floral elements: Coscinodiscus nodulifer, Nitzschia fossilis, N. marina, N. pandurformis, R. bergonii, and Thalassiosira oestrupii.

Correlation: The base of this zone approximates the calcareous nannofossil boundary NN18/NN19 as used by Backman (this volume). The *Pseudoeunotia doliolus* Zone correlates to the *Rhizosolenia curviostris, Actinocyclus oculatus*, and *Denticulopsis seminae* Zones of Barron (1980a).

EPOCH BOUNDARIES

Ryan et al. (1974) and Hailwood et al. (1979) place the middle Miocene/late Miocene boundary within the calcareous nannofossil *Discoaster hamatus* Zone (NN9) of Martini (1971) and correlate this epoch boundary with the lowest normal event of paleomagnetic Epoch 11. The middle Miocene/late Miocene boundary approximates the boundary between the *Rhizosolenia barboi* and the *Denticulopsis praedimorpha* zones as defined in this chapter.

The Miocene/Pliocene boundary is placed within the upper portion of the calcareous nannofossil Zone NN12 of Martini (1971) and is correlated to the middle of the lowest magnetically reversed interval of the Gilbert paleomagnetic Epoch by Ryan et al. (1974), Cita (1975), and Van Couvering et al. (1976). Burckle and Opdyke (1977) place the Miocene/Pliocene boundary at a level higher than the top of paleomagnetic Epoch 5 but lower than the "c" event of the Gilbert Epoch. This placement agrees with the previously mentioned workers and is based on analysis of the diatom flora from the "Tripoli" in the lower portion of the neostratotype Messinian Stage. The Miocene/Pliocene boundary falls in the upper portion of the *Thalassiosira convexa* Zone as used in this chapter and is approximated by the last occurrence of *T. miocenica* (Burckle, 1978b).

Burckle and Opdyke (1977) and Haq et al. (1977) place the Pliocene/Pleistocene boundary at the top of the Olduvai Event of the Matuyama paleomagnetic Epoch. Although others suggest that this placement should be slightly lower (see Ryan et al., 1974), the author adheres to the conclusions of Haq et al. (1977) and correlates the Pliocene/Pleistocene boundary with the top of the Olduvai. The Pliocene/Pleistocene boundary correlates with the lower portion of the *Pseudoeunotia doliolus* Zone as used in this chapter.

The paleomagnetic scale of Hailwood et al. (1979) is used in this chapter for the time interval from 11.0 to 65.0 m.y. For the younger interval of time (0–11.0 m.y.) the paleomagnetic scale of Ness et al. (1980) is adopted.

METHODS

Approximately 1.5 cm³ of sample was placed into a 400-ml beaker. The sample was disaggregated with the addition of 10 ml of 30% hydrogen peroxide. Upon completion of the resulting reaction, 30 ml of 37% hydrochloric acid was added. The sample was then gently heated until the liquid became light yellow in color. The sample's acidity was then neutralized by the addition of approximately 300 ml of distilled water, and then the sample was decanted after 2 hr. of settling. This decanting process was continued until pH 7 was reached. Strewn slides were prepared on 22 \times 40 mm cover glasses and mounted in Hyrax on 22 \times 75 mm glass slides.

Strewn slides of acid-cleaned material prepared aboard the *Glomar Challenger* were re-examined together with additional samples processed at onshore facilities. One cover glass was examined in its entirety at \times 500 with use of an Olympus BH-2³ light microscope. Species identifications were confirmed at \times 1250. Species were recorded as *abundant* if two or more specimen were present within one field of view at \times 500; *common* if one specimen occurred in two fields of view; *few* if one specimen was observed per horizontal traverse; and *rare* if specimens were encountered less frequently. Quality of fossil preservation of samples is based on the ratio of finely silicified taxa to robust and heavily silicified forms in each sample.

Samples within the uppermost 55 m of Hole 552A were examined for the fluctuation in diatom abundance, which is based on the average number of diatom frustules observed per field of view. A numerical value was obtained for each sample by dividing the total number of diatoms observed by the field of view encountered. At least 450 fields of view (0.5 mm) at a magnification of \times 500 were observed for each sample. The resulting numerical value for each sample was then placed into one of the following categories: Samples that are barren of diatoms, samples that average one diatom specimen observed per eight fields of view, samples in which at least one specimen was observed in two fields of view, samples that contain at least one diatom per field of view, and those samples in which two or more diatoms were observed for each field of view. The counting techniques of Schrader and Gersonde (1978) were employed during this study.

BIOSTRATIGRAPHY

Site 552

Site 552, located adjacent to DSDP Site 404 on the western margin of the Rockall Plateau, was drilled in hope of determining the chronology of the dipping reflector sequence present beneath the sediment cover. A secondary objective for Site 552 was to document the climatological and oceanographic changes within this region throughout the Tertiary Period.

Diatoms vary in abundance from rare to common in the Eocene to Quaternary sediments recovered from Site 552. Preservation is generally poor in the Eocene sediments, which have a flora dominated by *Stephanopyxis* grunowii, Melosira clavigeria, and Arachnoidiscus sp. Sponge spicules are common to abundant within the middle and uppermost Eocene sediments. Diatom preservation is usually moderate in the Neogene sediments. However, occasional intervals do occur where well-preserved and abundant diatoms are present.

Table 1 illustrates the zonal assignments of the cores recovered from Site 552 and other sites from Leg 81. It should be noted that a depth discrepancy of approximately 10 m exists between the depth of a prominent marker horizon observed in cores from both Holes 552 and 552A. Although the cause of this discrepancy is uncertain, the probable cause may be a miscount in the drill barrel length during the washing operation at Hole 552.

Hole 552

Twenty-five cores (79.19 m) were recovered from Hole 552 (56°02.56' N. 23°13.88' W. 2311 m water depth) by rotary drilling. A dissolution interval occurs in Cores 552-1 and 552-2. This interval is approximately 10 m stratigraphically higher in Hole 552 than in Hole 552A. Ice-rafted detritus is common throughout this interval. Sample 552-2, CC is Pliocene in age based on the occurrence of Thalassiosira convexa var. aspinosa, and T. oestrupii. The base of the Thalassiosira convexa Zone in Section 2 of Core 4 coincides with a dissolution interval that is recognized at all sites (see Fig. 10). In Hole 552 this dissolution interval occurs from Section 2 of Core 4 through Core 9. All samples examined within this interval, with the exception of Sample 552-6-3, 106-107 cm, either contain rare to few poorly preserved diatoms or are barren. Sample 552-6-3, 106-107 cm contains few moderately preserved diatoms and is tentatively placed in the Nitzschia porteri-Nitzschia miocenica Interval. This age assignment is based on the presence of N. reinholdii, N. marina, and T. eccentrica. The absence of T. convexa and Coscinodiscus yabei support this assignment.

One specimen of *Triceratium castelliferum* was observed in Sample 552-8, CC. This species is reported from the middle Eocene of the South Atlantic (DSDP Site 356; Fenner, 1977) and the late Eocene of Oamaru, New Zealand (Schmidt et al., 1874). The occurrence of this Eocene species suggests that a hiatus exists between the

³ Any use of trade names is for descriptive purposes and does not imply endorsement by the U.S. Geological Survey (USGS).

			<u></u>			Hole	e			
Age Pleist. Pliocene early late Miocene middle 1. Olig/e. Mio 1./m. Eocene e. Eocene	Diatom zone	552	552A	553	553A	553B	554	554A	555	
Age Pleist. Pliocene early late Miocene middle 1. Olig/e. Mio 1./m. Eocene	Pseudoeunotia doliolus		1-2 to 5-1?	1,CC		1,CC	1,CC			
	late	Nitzschia marina		8-3? to 10-2			2,CC			
Age Pleiss Pliocene Miocene 1. Olig/e. 1./m. Ec e. Ecc		Nitzschia jouseae		10-2 to 16-4				5,CC		
	early	Thalassiosira convexa	2,CC to 4-2	16,CC to 28,CC		1-2 to 2-2		7-3 to 8,CC		5,CC to 6-4
Ai Ple Pliocene Miocene	late	Nitzschia porteri Nitzschia miocenica Coscinodiscus yabei Denticulopsis praedimorpha Rhizosolenia barboi	6-3	29,CC to 30,CC					1-1 to 1,CC 3-4	6,CC to 9-1 9-2 to 11,CC 12-2 to 16-3 16,CC to 18,CC
	middle	Denticulopsis nicobarica Coscinodiscus lewisianus "Actinocyclus" ingens		36-3		7,CC to 8-4				19-1 to 21-2 21,CC
l. Olig/e. Mio		"Rocella" gelida				8,CC to 9-4				
		Unzoned	8,CC to 12-1	37-3 to 38-2		9,CC to 10,CC				
Pliocene early late Miocene middle 1. Olig/e. Mio 1./m. Eocene e. Eocene	Unzoned	12,CC to 21-3			11-4 to 21-1					

Table 1. Diatom zonation of samples from DSDP Leg 81 sites.

middle Miocene (Sample 552-6-3, 106-107 cm) and the Eocene (552-8, CC). This is in agreement with the nannofossil stratigraphy of Backman (this volume), in which a hiatus of approximately 25 m.y. occurs in Section 4 of Core 8 and separates the middle Miocene from the middle Eocene.

Few moderately preserved Eocene diatoms are present in Sample 552-10, CC. The presence of *T. castelliferum* and *Stephenopyxis grunowii* suggests a middle to late Eocene age. The absence of *Pyxilla reticulata, Trinacria pileolus, T. simulacrum*, and *T. excavata* var. *tetragona*, which are common within sediments assigned by nannofossil control to the early Eocene (see Hole 553A), supports this age call. The common occurrence of several species of *Arachnoidiscus* within this sample suggests a shelf environment.

Except for Cores 12, 14, and 21 (Core 11 had no recovery) all other samples examined contain poorly preserved specimens or are barren of diatoms. Cores 12, 14, and 21 are early Eocene in age based on the occurrence of *P. reticulata*, *T. pileolus*, *T. excavata* var. *tetragona*, and *Rhizosolenia interposita*. Numerous varieties of *Stephanopyxis* are common in Sample 552-14-2, 125-126 cm.

Hole 552A

Hole 552A (56°02.56'N, 23°13.88'W, 2311 m water depth) was hydraulic piston cored (HPC) from 0 to 183.5 m sub-bottom. Eocene and middle Miocene to Holocene diatoms are present throughout the 183.5 m of sediment recovered. Table 2 shows the stratigraphic occurrence of selected diatoms. Figure 5 shows the ranges of selected stratigraphically useful species. Reworking of Eocene, early Miocene, and middle Miocene species occurs sporadically throughout the hole.

Cores 1 and 2 are assigned to the *Pseudoeunotia doliolus* Zone and contain a well-preserved diatom assemblage that includes such species as *P. doliolus, Rhizosolenia bergonii, Hemidiscus cuneiformis, Thalassiosira oestrupii*, and *Nitzschia panduriformis.* The presence of *Rhizosolenia curvirostris* in Sample 552A-2-2, 52 cm through Sample 552A-2-3, 1 cm suggests that these samples have an age between 0.35 and approximately 1.5 m.y.

The placement of the base of the *Pseudoeunotia doliolus* Zone, defined by the first occurrence of *P. doliolus*, is difficult to determine because an interval of dissolution exists between Samples 552A-2-3, 30 cm and 552A-7-3, 119 cm. Samples within this interval are generally barren of diatoms. However, Sample 552A-5-1, 90 cm contains common well-preserved diatoms. The presence of *P. doliolus* in this sample places the base of the *Pseudoeunotia doliolus* Zone between this sample and Sample 552A-8-3, 91-92 cm, which is directly below the dissolution interval.

The presence of N. reinholdii, N. fossilis, and the silicoflagellate Mesocena quadrangula in Sample 552A-5-1, 90 cm, suggests that this sample is older than 0.79 m.y. ago, the age cited by Burckle (1977) for the last occurrence of M. quadrangula in the central Pacific.

The presence of N. fossilis, which has a last occurrence that approximates the last occurrence of M. quadrangula (see Barron, 1980a), supports this age for Sample 552A-5-1, 90 cm. Within the central Pacific, Koizumi and Kanaya (1976) place the last occurrence of N. fossilis between the Jaramillo Event of the Matuyama Epoch and the earliest part of the Brunhes Epoch.

Table 2. Stratigraphic occurrence of selected diatoms in Hole 552A.

Zone	Core-Section (interval in cm)	Abundance Preservation	Actinocyclus curvatulus A. divisus	A. ehrenbergii A. ellipticus A. ellipticus var. elonzota	A. ingens	Actinoptychus undulatus Arachnoidiscus sp.	Asterolampra marylandica	Asteromphalus elegans A. hookeri	Bacteriastrum hyalinum	Brightwellia cf. coronata	Cocconets vitrea Coscinodiscus marginatus	C. nodulifer C. nodulifer var. cyclopsis	C. occulus-iridis	C. radiatus C. stellaris	C. tabularis Denticularite buoline	Diploneis sp.	Ethmodiscus rex Conjothectum en	Hemiaulus polycistinorum	H. polymorphus	Mederia splendida	Melosina architecturalis	M. sulcata	Navicula lyra N. scutellum	Navicula sp.	Nittsenia cyimanca N. fossilis	N. jouseae N. marina	N. panderiformis	N. reinholdii Nitzschia sp.	Podosira glacialis	Pseudoeunotia doliolus Pterotheca aculeifera	P. danica P. cnada	Rhaphoneis amphiceros R. suirella	Rhizosolenia barboi	R. bergonii R. curvatulus	R. hebata R. praebergonii	r. stytorma Rouxia californica	Opephora sp. S. grunowii	S. turris Svnedra ulna	Thatassionema nitzschoides	I hatassiosing convexa val. азрипози Т. сопvexa var. сопvexa T. eccentrica	T. Jacksonii	T. leptopus T. miocenica	T. plicata T. pricata	T. symbolophorous T. symbolophorous	Thalassiosira sp. Thalassiothrir langissima	Triceratium cinnamomeum	Prinacria excuvata Xathiopyxis ovalis
	1-2, 30-33 cm	FM	RR							-	- R							-	- 1	2 -	- 1	2 -					R		- 1	R -				R -	R - 1	2 -			R -	R	- F	R			- R		
Pseudo- eunotia doliolus	1,CC	FM	— R						- R	-	- R	R -							- 1	- 1						- R	-			R -			-		R				R -	R	- F	R — F	2				
	2-3, 30-31 cm	B								-												17					-							27	R												
	2,00	AG	- R	R					- R		- R	R -		- R	R -		R -	-	- 1	- 1		- R				- F	-		- 1	F -			-	RR	R			RR	F -	R	- B	x			- C		
	3-1, 63-66 cm	B _	- R							_												2							_																		
	4-1, 84-85 cm	в —					_			_			_			_									_		_						_	:		_											_
	4,CC	в —					-			-			-														-						-														-
	5-1, 36-37 cm	RP						= -		-								-			_						-																				
	5-2, 15-16 cm	в —								-															-		-		·				-						R -								
	5,CC	в —							• =	-			-											-			-																				
	6-1, 94-95 cm	RP							-	-	- R																-																				
	5,CC	B				10				-								-						-			-				1		-														
	7.CC	B _								_																																					
Nitzschia	8-1, 90-91 cm	B -								-														_			_						-														
marina	8-3, 91-92 cm	RP								-	- R										- 1	2 -					-						_														
	9-1, 113-114 cm	CM	RR	R R -				- F	2 -	-	- F	R -		R —	R -							- R			- F	- F	-	R -	- R -				-	R -				- R	A -	C	- F	2 - F			RF		- R
	9-3, 53-54 cm	AM	- R	R							- R			- R			R -		- 1	- 1		- R			- R		-	R -					-	R	R				C -	F	- R	4 - R			- F		R
	9-4, 13-14 cm	AG	RR	R					17	-	- R						R -		- 1	- 1					- R	- R	-	R -				R -	-					- R	C -	F	- 8	k - F			RF		
	10-1 16-17 cm	AG	PP	RR		_			2 0	-	- r	E _	_	- P		P	P _		- :	5 -		P			P	- K		R -	_				K	R _	P			P P	CI	D D D		$\dot{e} = \ddot{e}$		P	PF		
	10-1, 42-43 cm	CM	- F	8	_	_				_	- F	8 -				-	R -		- :	1-		- 8		=1.	- 8	- 8		RR					_	8 -	R			RR	F -	- R F	- 1	8 - F			RF		_
	10-2 34-35 cm	A G	DD	P	1			_	P	2005	P	P	1.2.1	- 0	P		P		- 1	7			P	P	-	D D	_	D D		2002	-	_ p	_	<u> </u>	P	-	P	FP	CI	FFF	- 1	F - T		1	PF	_	P
	10-3, 61-62 cm	AG	_ R	RR-		R -	_	R -	- R	_	- R	F -		- R	R -	- R	R -		- 6			- R	8 -	213	- R	RR		FR	R -				R		R		R -	RF	CI	FRF	RI	$\hat{e} = \hat{r}$		R -	RC		
	10.CC	CM	- R	RR-				- 1	RR	_	- R	R -	_			- R			- 1			- R	R -		- R	RR	-	F				- R	-	R -	R			R -	FF	R - R	- F	R - F	t		- F		
	11-1, 83-84 cm	FM	RR	R						-	- R	R -							- 1	- 1						RR	-	R -	-				R		R			R -	RF	X R R	- F	8 — F	t		- R		
	11-3, 23-24 cm	AM	- F	R						-	- R	R -					R -	-	- 1			- R			- R	CR	-	FR				- R	C		R — -			R -	FF	AR F	- F	f - R		- R -	- F		
	11,CC	CM	- R	777					- R	-	- R	R -	-			- R	R -		- 1	-		- R	R —		- R	RR	-	FR				- R	R					R -	CB	x - R	- B	K - B		- R -	RF		
	12-3, 47-48 cm	CG	D D	8		_			P			P _		P		P			- 1					P	P	PP	_	P P	-			_ R	R		P			RR	RI	2 _ R				P	RR		
Nitzschia	12.CC	AG	- R	R					RR	-	- R	F -					22		Ξ;			- R	22	R		RR	_	RR	Ξ.			- R	F	R -	R - 1	RR		R -	C I	R - R	- F	R - F	R -	- R -	- 0		
jouseae	13,CC	CM	RR	R					- R	-	- R	R -				- R	R -		- 1	- 7						RR	-	RR				- R	R		R		R -	RR	F -	R	- 1	R - F		- R -	- R		
	14-1, 100-101 cm	CG	— F	R						-	- R	R -		— R	R -	- R			- 1	- 1		- R	R —		- R	RR	-	RR				- R	F		R			RR	FF	RRR	RF	A — F			RF		
	14-3, 100-101 cm	AM	- F	- F -						-	- F	C -			R -				- 1	- 1				-1-	- R	FR	-	FR				- R	R					R -	CF	k - R	- P	x - R			RF		
	14,00	FM	- R	p						-	- R	R -							- 1	-					- R	P P	-	R -				- K	R		p		p	p 0	K -	R	- 8	K - R		P	P P		
	15-5, 100-101 cm	AG	A R	n - 1	1	_		_ 2	P	1	- K	E			K -	R			- 1	-		R			F	FF		FF	_			R	F		R		R	R -	A -	R	RI	$\hat{R} = r$			R K		
_	16-1, 130-131 cm	AG	- R		2 -	R -		- 1	RR	-	- R	CR	R	- R		-	22	12	- 1				R -	R -	- R	- R	_	FR					F		R		R -		A -	R	- 1	R - F			- A		_
	16-4, 7-8 cm	CG	- R			R -				-	- F	RR				- R			- 1	- 1		- R				RR	-	R -	-		- r	- R	R		R		R -	- R	FF	1 - R	- F	R - F			- R		

	16.CC	F M - R R	L R I	R R R R	R	R	- R	- R - R -	F R	R R	R	- R	R R R	R - R	R
	17-1.66-67 cm	CG-RR-R		R R F	R	R R -		R C - R -	F F	R R	R	-R-RR	FR-RR	F - F	$\mathbf{R} = -\mathbf{R} = -\mathbf{R}$
	17.2 64 66	E D D				D		D D	D	D D			E D	p	
	17-2, 04-00 cm	F F - K				K		A -	A	A			$r = - \kappa - $	n	
	17,00	F P - K		R R		K		K K -		K			$\kappa \kappa -$	к - к	K
	18-3, 83-85 cm	F P F		R		R		- R		R	R		R R -	R	R
	18,CC	R P r B	r - l	R	- 1	I						r R -	R R		
	19-1, 95-96 cm	R P r R		8	R	r r	- R					r R -		R	
	19.1 95-96 cm	AG-RFFR	- R 1	R R R	P	F P	_ P	FC-F-	R	R	R		C R - R -	R - R - R	B C
	10.00				x	n	- n p	DE D	n	D	D		E D D	D D	D D D D D
	19,00	CO-KFFK		N F N		A A -	n	R I - R -	n n	~	A	n . n	K K	A - A	
	20-1, 82-83 cm	F M = -R R = -R	(r - l	R R R	$ \mathbf{R} \mathbf{r} $	TTRRT	- R	K K - K -	R R T			- K F K -	к к -	K	K K
	20-2, 83-84 cm	C G = R F F = -F	- R H	R R R		R R -		F F - R -	R	R	R		C R R	R - R	R = R F =
	21-1, 124-125 cm	R P		8	R		- R						R		
	21-3, 64-65 cm	CG-RFF	R R H	R R R		F R -	- R	R R - F -		R	R		R R R	R - R	-R-R-R
	21.00	AG-RCRR	_ P P	F R		F	P _ P _	RE-R-	R	R			CRR	R	P P - P - P
	22 1 124 126 m	C C F P P		D D D	p p	E .	D A	FFP	F	D			E D E D	D	P P F
	22-1, 124-126 cm	C 0 - F K K		K K K	K - K		- K	$r r - \kappa -$	r	ĸ			F K - F K	R	$\mathbf{k} = \mathbf{k} \mathbf{r} = -$
22 57 17	22-2, 124-125 cm	K P				K K -	- R	R K -					$\mathbf{k} \mathbf{k} - \mathbf{k}$	K	R
Thalassi-	22,CC	CM RRFR- -R	- - - - - -	R R R	R - R	R	R-	R	R	R	R	R	F R - R -	R R	R F
osira	23-1, 20-21 cm	F M - R R R R	L - R - - R	- R R		R	- R	R R		R	R		FR - RR	R	R
convexa	23-2, 123-124 cm	R P - R		R R		R	- R	R -					R -		
Carles Portan	23-3, 34-35 cm	RMRRR		R	R	8	- R	R-				R -	R R -	R	
	23.00	EM-PPP		D		P		D		D	p	p	D D D	D D	P P
	24 1 47 48			E D	D -					n	x		R R - R -	n n	
	24-1, 4/-48 cm			K F K	K T	K K -	- K	<u> </u>	K	K		$- \kappa - \kappa -$	F K - K -	к к	$\mathbf{K} = -\mathbf{F} = -\mathbf{K}$
	24-2, 45-46 cm	C M K K K - K		CR - RR		R R -	- K	R F - R -	R	R	R	-R R	F = -FR	F R	R R -
	24,CC	CM - RRR - R	-R -	R R R R - R	R - R	R R -		F F - R -		R - R		R -	FR-RR	R R	R F
	25-2, 140-142 cm	A G - R C F R	- R	R = R		F R -	- R	F F - F -	F	R	R	R	CR-FF	R	F
	25-3, 40-41 cm	CMRRCFR	- R - R F	8 F F	R - R	R R -		R R - R -		R	R		C R R	R R	R F R
	25 CC	8 P		- R		R							P		
	26.1.70.71 cm	AM PPP		D D E	D	D D	p	D F D	E	D	D		C D D	p	D E D
	20-1, 70-71 cm	A M - A A A		A A I	x	n n -		K F - K -	r	K	x		CKK	K	$\kappa r\kappa$
	20-2, /0-/1 cm	K P		<pre></pre>									x		
	26,CC	B													
	27-1, 30-31 cm	FMRRRRR	- R F	R = R		R	- R	R	R	R		R -	F R - R R	R R	R
	27.CC	CMRRRRF	- R H	R R R		R	- R	R R -		R	R		C R - R -	R R	
	28.CC	FM-RRRF	- R F	R = R		R	- R	R R -	R	R	R		F R - R -	R R R	R R
							-								
	29-1, 137-138 cm	F M - R R R	F			R	R	R R -	R				F R -	R	R
	29,CC	F M - R R R F		R		R R -	- R	F R -	R				R R -	R R	
	30-2, 95-96 cm	F M R F		R		R R -		R			R		F R -		R
	30.CC	FM-RFRR		R		R	- R R	RR-F-	R		P		C R	R	F R
	31-1 45-46 cm	R P R							*				P		P
	21 CC	P P											N		K
	33,00	K F													
	32-2, 95-96 cm	F P =		K K		K					R		R		R -
	32,CC	R P					- R								
Nitzschia	33-1, 50-51 cm	R P		2								R -			
porteri-	33.CC	R P		2											
Nitzschia	34-2, 50-51 cm	B													
min	34.00	B								23222					
mile	35.00	P													
cenica	33,00	B													
	30-3, 30-32 cm	K P R -													
	36,CC	B													
	37-2, 34-35 cm	B													
	37-3, 99-100 cm	F M	- R									R			
	37.CC	R P	- R									R			
	38-1 121-122 cm	F M	B							R					
	38 2 31 32 ar	F M								P		D D			
	30-2, 21-22 cm									A		K K -			K -
	38,00	к м – – – – – – – –										K			

Note: R = rare, F = few, C = common, A = abundant, r = reworked.



Figure 5. Stratigraphic ranges of selected diatoms at Hole 552A.

The paleomagnetic results of Leg 81 (see Zimmerman et al., this volume) place the base of the Jaramillo Event between Samples 552A-5-1, 85 cm and 552A-5-1, 112 cm. The Matuyama Brunhes Epoch boundary is placed in Section 1 of Core 4. These data suggest that the presence of N. fossilis in Sample 552A-5-1, 90 cm is slightly below the species' last occurrence and that the true last occurrence of N. fossilis is not observed because of dissolution.

Other species present in Sample 552A-5-1, 90 cm include Denticulopsis seminae, D. seminae var. fossilis, and Roperia tesselata. These species are indicative of both higher latitudes (D. seminae and D. seminae var. fossilis) and lower latitudes (R. tesselata) and may represent the influx of colder polar waters into the temperate waters of the Rockall Region.

The *Nitzschia marina* Zone is partially represented by a dissolution interval that is dominated by ice-rafted detritus. This interval extends from Sample 552A-2-3, 30 cm through Sample 552A-7-3, 119 cm and can be correlated to a similar interval present at Hole 552.

Although the last common occurrence of T. convexa occurs at Sample 552A-10-2, 0-3 cm, the true last occurrence of this species is at Sample 552A-9-1, 63 cm. The paleomagnetic results of Leg 81 place this event a third of the way between the Gauss/Matuyama boundary and the Olduvai paleomagnetic Event. The placement of this datum agrees exactly with the results of Burckle and Trainer (1979).

The last occurrence of *N. jouseae*, which defines the base of the *Nitzschia marina* Zone, occurs between Sample 552A-10-2, 30 cm, and 10-2, 20 cm. The paleomagnetic results of Leg 81 place these samples approximately midway between the top of the Kaena Event and the Gauss/Matuyama boundary. The age of this datum event therefore agrees well with an age of 2.6 m.y. ago, assigned by Burckle and Trainer (1979) for this event within the central Pacific (see Fig. 6).

The only specimen observed of R. praebergonii appears in Sample 552A-10-1, 70-73 cm. The presence of this warm-water species, which is confined typically to the tropics (see Burckle, 1972, 1978b), within the Rock-all Region reflects the influence of the North Atlantic Drift within this region.

Koizumi (1968) records *R. praebergonii* in sediments examined from the Oga Peninsula of Japan. Barron (1980a) similarly records this species off the northeast coast of Japan. The presence of this species at high latitude is the result of its having been transported by the warm-water, northward-flowing Kuroshio Current, a situation similar to the North Atlantic Drift influence within the Rockall Region. *R. praebergonii* is also recorded by Schrader (1977) from DSDP Sites 332 and 333 within the central Atlantic.

The Nitzschia jouseae Zone is represented in the interval from Samples 552A-10-2, 30 cm to 552A-16-4, 7-8 cm. The base of this zone is defined by the first occurrence of N. jouseae.

Burckle (1972, 1978b) correlates the first occurrence of *N. jouseae* with the "C" Event of the Gilbert Epoch. Therefore this datum approximates 4.5 m.y. ago. Using this date for the first occurrence of *N. jouseae* in the North Atlantic results in a somewhat uniform sediment accumulation rate (see Fig. 7) which suggests that this datum event at Site 552 is isochronous with the equatorial Pacific.

However, the placement of the first occurrence of *N. jouseae* between Sample 552A-16-4, 7-8 cm and 552A-16,CC is in slight disagreement with the nannofossil results. Backman (this volume) places the calcareous nannofossil NN13/NN14 Zonal Boundary (which approximates 4.3 m.y. ago) between Samples 552A-16,CC and 552A-18,CC based on the occurrence of *Amaurolithus primus* and *Discoaster asymmetricus* in Sample 552A-16,CC (Zone NN14) and *Ceratolithus rugosus* in Sam-





ple 552A-18,CC (Zone NN13). The occurrence of C. rugosus in Sample 552A-18,CC suggests that the stratigraphic first occurrence of N. jouseae should be slightly lower.

The last occurrence of N. cylindrica occurs between Sample 552A-17-1, 66-67 cm and 552A-16, CC. Burckle (1978b) places this datum above the "C" Event of the Gilbert Epoch. The first occurrence of N. jouseae is placed within the upper portion of the "C" Event of the Gilbert Epoch. If the placement of the last occurrence of N. cylindrica is correct, then the last occurrence of N. jouseae should be stratigraphically lower. This would also suggest that this datum is not isochronous, although



Figure 7. Sediment accumulation rate curve calculated for Hole 552A from diatom datum levels.

this would agree better with the nannofossil data. The reliability of N. cylindrica is questionable (see Burckle, 1978b), whereas the isochroneity of the first occurrence of N. jouseae between the North Pacific and the Atlantic, as warranted by the sedimentation curve, suggests that the latter datum is more consistent and reliable. Burckle (1978b) suggests that the last appearance of N. cylindrica in high latitudes may be diachronous. Furthermore, a rank and reliability factor of third order was assigned to this datum based on the difficulty in identification and the numerous local ecological or geographical restrictions. Contrary to this, the first occurrence of N. jouseae is assigned a first-order ranking (Burckle, 1978b) based on the ease of identification and "common and persistent either first rapid evolutionary appearance or highly consistent range." This suggests that this occurrence of N. jouseae is the true first occurrence. However, further quantitative studies are required to resolve the disagreement between diatoms and nannofossils.

The base of the *Thalassiosira convexa* Zone is defined by the first occurrence of *T. convexa*. A secondary marker that is nearly synchronous with the *T. convexa* event is the first occurrence of *T. miocenica*. Both species first appear in Sample 552A-28, CC. The first occurrence of *T. oestrupii* is in Sample 552A-21-3, 64-64 cm. This datum is placed by Burckle and Opdyke (1977) within the lowest portion of the Gilbert Epoch and is suggested by Barron (1980a) to approximate the Miocene/Pliocene Boundary of Van Couvering et al. (1976).

The last occurrence of *T. miocenica* that also approximates the Miocene/Pliocene boundary is in Sample 552A-22, CC. Burckle (1978b) correlates this event to midway between the "C" Event of the Gilbert Epoch and the top of Epoch 5. *T. praeconvexa*, which has a very short stratigraphic range, is correlated by Burckle (1978b) to occur from the middle of Epoch 6 to the middle of the lowest normal event of Epoch 5. Within DSDP Hole 552A, *T. praeconvexa* occur from Samples 552A-32-2,

95–96 cm to Sample 552A-28, CC. The range of this species slightly overlaps the basal boundary of the *Thalassiosira convexa* Zone and is a useful secondary marker.

The Nitzschia porteri-Nitzschia miocenica Interval is recognized from Sample 552A-32-2, 95-96 cm to Sample 552A-28,CC based on the occurrence of *T. praecon*vexa below the first occurrence of *T. miocenica* and *T.* convexa. N. aff. miocenica is also observed within this interval.

The abrupt downcore change in diatom preservation observed at all sites (see Fig. 10) at approximately 6.2 m.y. ago occurs within the uppermost portion of Core 31. The presence of *Actinocyclus ingens* in Sample 552A-36-3, 30-32 cm suggests that this sample is middle Miocene in age. Section 3 of Core 37 through Core 38 is Eocene in age and contains an assemblage similar to that found in Cores 552-10 and 552-12. Sponge spicules are common within this interval.

Site 553

Three holes were rotary cored at Site 553, located northwest of Site 552 on the Hatton Drift, which extends along the western margin of the Rockall Plateau. Preservation of diatoms in the Eocene to Recent sediments recovered is similar to that at Site 552. Preservation is generally moderate, however, two dissolution intervals occur: one is Pliocene-Pleistocene and the other is late Miocene in age. The Pliocene-Pleistocene interval is dominated by ice-rafted detritus. Both dissolution intervals are equivalent in age to similar intervals recognized at Site 552.

A single mudline core taken at Hole 553 contains well-preserved Pleistocene diatoms which are assigned to the *Pseudoeunotia doliolus* Zone based on the presence of *P. doliolus*.

Hole 553A

Fifty-nine cores (531.5 m) were recovered from Hole 553A (56°05.32'N, 23°20'W, 2335 m water depth). A reentry cone set with casing was placed at a subsurface depth of 56.5 m, resulting in a late Miocene-early Pliocene age for both Cores 1 and 2. These cores are assigned to the *Thalassiosira convexa* Zone. The Miocene/Pliocene boundary approximates the first occurrence of *T. oestrupii* in Sample 553A-1,CC.

The interval of dissolution present at the base of the Thalassiosira convexa Zone throughout the Leg 81 sites occurs from Core 2 through Section 3 of Core 7. Actinocyclus ingens and Denticulopsis hyalina are both observed in Sample 553A-7, CC and occur stratigraphically down to Sample 553A-8-3, 103 cm, indicating a middle Miocene age for the interval from Sample 553A-7,CC to Sample 553A-8-3, 103 cm. A hiatus occurs directly below this interval. Diatoms are very rare from Section 4 of Core 8 through Section 3 of Core 9. However, this interval is earliest Miocene in age based on the rare occurrence of Rocella gelida. The frequence of R. gelida increases downcore, becoming common in Section 3 of Core 9. Weaver and Gombos (1981) place the Oligocene/Miocene boundary within the range of Bogorovia veniamini just above the acme of R. gemma (this species This would suggest that the Oligocene/Miocene boundary change of La Brecque et al. (1977) can be approximately placed in Section 3 of Core 9 (94-95 cm). However, diatoms are rare within the interval and no secondary markers are available. Nannofossil control (Backman, this volume) records the extinction of *Dictyococcites bisectus*, which approximates the Oligocene/Miocene boundary of Hailwood et al. (1979) between Samples 553A-9-4, 5 cm and 553A-9-4, 15 cm, therefore suggesting a slightly lower stratigraphic placement of this boundary.

A third hiatus is recognized between Samples 553A-9-4, 67-68 cm (late Oligocene) and Sample 553A-9,CC, which is assigned to the Eocene. An Eocene age is also assigned to samples examined in Cores 10 and 11. The presence of *Coscinodiscus oblongus*, *Brightwellia coronata*, and rare *Trinacria* species in Core 10 suggests a middle to late Eocene age. The presence of common *Trinacria* [*T. pileolus*, *T. excavata*, *T. simulacrum*, and *T. excavata* var. *tetragona*], *Peterotheca aculifera*, *Stictodiscus angulatus*, *Pyxilla renticulata*, *Rhizosolenia interposita*, and *Sceptroneis gemmata* in Core 11 suggests a late Paleocene to early Eocene age. Locally these species are common throughout the interval assigned by nannofossil control (Backman, this volume) to the early Eocene.

All samples examined below Core 11 are barren of diatoms, with the exception of Samples 553A-18,CC, 533A-21,CC, 553A-22,CC, 553A-23,CC, 533A-24,CC, and 553A-37-2, 28 cm. Samples 553A-18,CC, 553A-21,CC, 553A-22,CC, and 553A-23,CC contain nondiagnostic diatom frustules in which silica is replaced by calcite. Samples 553A-24,CC and 553A-37-2, 28 cm contain pyritized diatom frustules of *Triceratium* sp. and *Coscinodiscus* sp.

Hole 553B

Of the four cores recovered from Hole 553B (56° 05.32'N, 23°20.61'W, 2399 m water depth) only Core 1 contains moderately preserved diatoms. This core is assigned to the *Pseudoeunotia doliolus* Zone. Cores 2 through 4 contain either nondiagnostic specimens or are barren of diatoms. Ice-rafted detritus is present throughout these cores. This interval correlates to a similar interval of dissolution present within the *Nitzschia marina* Zone at Site 552.

Site 554

Site 554, located slightly northwest of Sites 552 and 553, was rotary cored to examine the nature of the outer high which forms the western boundary of a suite of dipping reflectors (see Roberts, this volume). Diatoms are generally poor to moderately preserved within the sediments recovered.

Hole 554

Eight cores (76 m) of upper Miocene to Holocene sediments were recovered from Hole 554 (56°17.41'N, 23° 31.69'W, 2584 m water depth). Diatoms are poor to moderately preserved. Samples examined from Core 1 are assigned to the Pleistocene *Pseudoeunotia doliolus*. Zone based on the presence of *P. doliolus*. The occurrence of *Rhizosolenia curvirostris* in Sample 554-1,CC suggests that this sample has an age between 0.35 and 1.5 m.y.

The Pliocene–Pleistocene dissolution interval observed at both Sites 552 and 553 occurs within Cores 2-6 of Hole 554. With the exception of Sample 554-5,CC, all samples examined from this interval are either barren or contain rare nondiagnostic specimens. Sample 554-5,CC is tentatively assigned to the *Nitzschia jouseae* Zone based on the very rare occurrence of *N. jouseae*. Other species present include *Thalassiosira oestrupii*, *Nitzschia fossilis*, *N. reinholdii*, *Coscinodiscus nodulifer*, and *R. barboi*.

Cores 7 and 8 are placed in the late Miocene portion of the *Thalassiosira convexa* Zone based on the presence of *T. convexa*, *T. miocenica*, and *Thalassiosira* sp. 1. The occurrence of *T. praeconvexa* in Sample 554-8,CC suggests that this sample approximates the base of the *Thalassiosira convexa* Zone.

Hole 554A

Hole 554 was abandoned because of deteriorating weather conditions. After approximately a 5-hr. delay, weather conditions improved enough for drilling operations of Hole 554A ($56^{\circ}17.4'N$, $23^{\circ}31.65'W$, 2584 m water depth) to commence. The top 76 m of sediment was washed down to reach the approximate level where drilling was stopped at Hole 554. Six cores (137 m) of lower Eocene to upper Miocene sediments were recovered. Diatom preservation throughout this hole is generally poor although several samples were observed that contain common moderately preserved diatoms.

Sample 554A-1-1, 15-17 cm is placed in the uppermost portion of the Nitzschia miocenica Zone of Burckle (1972). This age assignment is based on the presence of Thalassiosira praeconvexa below the first occurrence of T. convexa and T. miocenica within Sample 554-8.CC. Therefore the base of the Thalassiosira convexa Zone, defined by the first appearance of T. convexa, occurs between Sample 554-8, CC and 554A-1-1, 15-17 cm. Burckle and Trainer (1979) correlate the first occurrence of these datums with the upper portion of Epoch 6, which suggests that these samples have an age of approximately 6.2 m.y. Nannofossil biostratigraphy places the NN11/NN12 boundary (approximately 6.0 m.y.) between Sample 554A-6-5, 46 cm and Sample 554A-6, CC. The calcareous nannofossil Zone NN11 extends from Sample 554-6,CC through Sample 554A-1,CC. This sample interval is assigned by Backman (this volume) to the Discoaster neorectus subzone of Bukry (1973) which correlates with the lowest portion of Zone NN11 (approximately 9.6-10.3 m.y.). Although this discrepancy between diatom and nannofossil biostratigraphy is present, further work is required to explain what factors are responsible. Radiolarian biostratigraphy (Westberg-Smith and Riedel, this volume) for Samples 554-8,CC and 554A-1-1, 18-20 cm suggests that these samples are near the evolutionary transition between S. delmontensis and S. peregrina and therefore approximate the S.

perigerna-D. pentultima Zonal boundary (approximately 6.5 m.y.).

Sample 554A-1,CC contains few moderately preserved diatoms. However, no age diagnostic forms were observed. The presence of numerous varieties of the genus *Nitzschia* suggests that this sample is younger than 9.0 m.y.

Sample 554A-3-4, 90-92 cm is tentatively placed within the *Coscinodiscus yabei* Zone. This zonal assignment is based on the presence of *Hemidiscus cuneiformis*, which has a first occurrence at approximately 12.0 m.y. ago, and the absence of the numerous *Nitzschia* species, which have an approximate first occurrence at 9.0 m.y. ago. Supporting this zonal assignment is the presence of *Thalassiosira* sp. 1 of Barron (1980a). All other samples examined from Hole 554A contain no age-diagnostic specimens or are barren of diatoms.

Site 555

Site 555 is located northeast of Site 554 within the shallow water of Hatton Bank. This location was selected in an effort to complete the dipping reflector transect from oceanic to continental crust. Furthermore, plans were made to HPC the Pliocene–Pleistocene sediments within this region for paleoceanographic reconstruction. However, time restrictions permitted only the rotary coring of a single hole.

Hole 555

Ninety-eight cores (505 m) of upper Paleocene to Pleistocene sediments were recovered from Hole 555 (56° 33.70'N, 20°46.93'W, 1669 m water depth). No siliceous upper Pliocene or Pleistocene sediments are present. Diatoms are generally rare to abundant throughout the upper 28 cores recovered. However, within this interval occasional samples are barren of diatoms. Generally samples examined stratigraphically below Core 22 are barren of siliceous material. Diatoms are especially common throughout several intervals within middle Miocene sediments. Table 3 shows the stratigraphic occurrence of selected diatom species. The ranges of selected stratigraphically important species are recorded in Figure 8.

Samples examined from Cores 555-1 through 555-4 are either barren or contain no age-diagnostic species. Species observed include *Hemidiscus cuneiformis*, *Actinoptychus undulatus*, *Thalassiosira eccentrica*, and *Nitzschia reinholdii*.

Common moderately preserved diatoms are observed in Core 555-5 through Section 555-6-4. This late Miocene interval is assigned to the *Thalassiosira convexa* Zone based on the presence of *Thalassiosira convexa* s. ampl., *T. miocenica*, and *Thalassiosira* sp. 1. The interval of dissolution presence at the base of this zone at all other Leg 81 sites occurs from Core 555-6 through Section 555-8-4.

Sample 555-8,CC contains N. cylindrica, H. cuneiformis, N. marina-reinholdii, and Rhizosolenia barboi. This sample is assigned to the Nitzschia porteri-Nitzschia miocenica Interval.

The Coscinodiscus yabei Zone is assigned to the interval from Section 555-9-2 through Section 555-11-2 based on the occurrence of *C. plicatus, Denticulopsis*

Zone	Core-Section (interval in cm)	Abundance Preservation Actinocyclus curvatulus A. divisus A. ethrenbergii A. ethroires	A. ingens A. ingens Actinopytchus undulatus Asterolampra marylandica Coscinodiscus lewisianus C. marginatus	C. noduitjer C. obsurus C. otulus-tridis C. pilicatus C. sadiatus C. saliaburyanus C. saellaris C. tabularis C. vetustissimus	C. yabei Craspedodiscus coscinodiscus Cussia paleaceus Denticulopsis dimorpha D. hustedtii D. hustedtii	D. lauta D. lauta D. praedimorpha D. pructata D. prunctata var. hustediti	Ethmodicus rex Hemiaulus polymorphus Hemidiscus cuneiformis Hyalodiscus dentatus Marrona stella	Mediaria splendida Melosina clavigena M. sulcata Nitzschia cylindrica N. fossilis	N. Jossils/N. reinholdii N. marina N. praereinholdii N. reinholdii Opephora sp.	Rhizosolenia barboi R. hebatata R. miocenica R. praebarboi R. styliformis	Rouxia californica Stephanopyxis turris Stephanopyxis sp. Synedra jouseae S. miocenica Thalassionema nitzschoides Thalassionema nitzschoides Thalassionera nitzschoides Thalassionera nitzschoides Thalassionera nitzechoides Theorenica T. miocenica T. praeconvexa Thalassiosira sp.	Thalassiothrix longissima Triceratium cinnamomeum
Thalassiosira convexa	1,CC 2,CC 3,CC 4,CC 5-5, 55-56 cm 5,CC 6-4, 76-77 cm	B		R			R			R R R		R - R - C - R - R - R - R - R -
Nitzschia porteri- Nitzschia miocenica	6,CC 7-4, 71-73 cm 7-6, 23-24 cm 7,CC 8-3, 70-71 cm 8-4, 63-64 cm 8-5, 20-21 cm 8,CC 9-1, 66-68 cm	R P R M R R R M R R R C M R R F R F P R R F P R R F M R R R C M R R R R T N N R R R R 1 R P R P -		R R			R	R R R R R R R R R R R R R R R R R R R R R R R R		R		F
Coscinodiscus yabei	9-2, 80-82 cm 9,CC 10-1, 128-129 cm 10-2, 67-68 cm 10-4, 30-32 cm 10,CC 11-2, 52-53 cm 11,CC	F M R - B - - B - - B - - B - - B - - B - - B - - F M R R R - F M R R - -	- R - C	R R R R R			- R R			R	R - R R	R — R — R R R —
Denticulopsis praedimorpha	12-2, 62-63 cm 12,CC 13-1, 15-16 cm 13;CC 14-1, 15-16 cm 14,CC 15-1, 75-76 cm 15,CC 16-3, 119-120 cm	C G R — R — R P — R R C G R — R R R - R M — F M R — R R - F M R — R R - F M R — R F M R F M F M F M F M F M F M F M F M	- R R - R - F R - R - R R R		R	R F	R R R 	R — R — — — — R — — — — — R — — — — — R — — — R — R —	R - R R R R 	R R R R F R R R R R R R R R R R R R		R R R R R R R R R R R R R R R R R R R
Rhizosolenia barboi	16,CC 17-3, 63-64 cm 17-6, 57-58 cm 17,CC 18-2, 38-39 cm 18,CC	F M R R R F P R F M R R - F M R R - C M R R -	- R R - R R - R - R R - R F - R R			R - R R - R R - R R		R — R — — R — R — — R — R — — R — R — — R — R —		R — — R R R R — — R — R R — R — R — R —		R — R — R — R — F —
Denticulopsis nicobarica	19-1, 76-77 cm 19-2, 42-43 cm 19-3, 37-38 cm 19-3, 37-38 cm 20-1, 87-88 cm 20,CC 21-1, 127-128 cm 21-2, 121-122 cm	C G R - R R I C G R R - A G R - R R - F M R R - R P R R - F P R R - C M R R -	R R R — R - F F — — R - R R — R R - R R — — R - R — — — — - R R — — — - R R — — — - R R R — R	R - R R R R R R R - - - R - R - R - R - - R - R - R - - - R R R R R - - - - - R - - R - - R - - - R R - - R - - - - - - - - - R - - R - <td< td=""><td>F R R F - R R F - - R - C - - - - F - - - - F - - - R - F - - - R R - - F - - - R - - R - - R R - - F -<</td><td>- R R R F R R R - R R R R - R R - R R - R R R R R R R R R R - R R R - R R</td><td>R R R R - R R </td><td>R — R — — R — R — — - — R — — - — R — — R — R — — - — R — — - — - —</td><td></td><td></td><td>R R R R F R R R R R F R R R R R C R R R R R R R R R <</td><td>A C F R F F F</td></td<>	F R R F - R R F - - R - C - - - - F - - - - F - - - R - F - - - R R - - F - - - R - - R - - R R - - F -<	- R R R F R R R - R R R R - R R - R R - R R R R R R R R R R - R R R - R R	R R R R - R R 	R — R — — R — R — — - — R — — - — R — — R — R — — - — R — — - — - —			R R R R F R R R R R F R R R R R C R R R R R R R R R <	A C F R F F F
C. Iewisianus	21,CC 22-1, 106-107 cm 22-2, 6-7 cm 22,CC	F M — R R - F P — R P — R P	- F R R R R - - F R - R R - - R	- R - R R R - R R 	R R F - - R	R — R R — — — — — — — — — — — — — — — —	R R	R — R — — R — R — — — — — — — —		- R R R F R R	R R R R - R R	C R

Table 3. Stratigraphic occurrence of selected diatoms in Hole 555."

Note: R = rare, F = few, C = common, A = abundant, r = reworked.



Figure 8. Stratigraphic ranges of selected diatoms from Hole 555.

hustedtii, and *H. cuneiformis*. Preservation within this interval is poor to moderate.

Few to common, moderately to well-preserved diatoms occur in Cores 12 through 16. Species present within this interval include *D. hustedtii*, *D. punctata*, *D. praedimorpha*, *D. hyalina*, *C. yabei*, *C. plicatus*, *Actinocyclus ingens*, and *H. cuneiformis*. This late middle Miocene assemblage is assigned to the *Denticulopsis praedimorpha* Zone.

The Rhizosolenia barboi Zone occurs in Cores 17 and 18. The base of this zone is defined by the first occurrence of R. barboi, which occurs in Sample 555-18,CC. Other species present include D. punctata var. hustedtii, R. praebarboi, A. ingens, and A. ellipticus.

Cores 19 through 21 are assigned to the *Denticulopsis* nicobarica Zone based on the presence of *D. nicobarica* and *D. punctata* var. hustedtii below the first occurrence of *R. barboi*. Additional species present include Asterolampra marylandica, *R. miocenica*, Craspedodiscus coscinodiscus, and Actinocyclus ingens.

Rare specimens of *C. lewisianus* are found in Samples 555-21,CC and 555-22-1, 106 cm. Therefore these

samples are placed within the *Coscinodiscus lewisianus* Interval. The remaining portion of Core 22 is poorly preserved, and only specimens of *A. ingens* are observed.

All other samples below Core 22 with the following exceptions are barren of diatoms. Core 27 contains fragments of the Eocene diatom *Stephanopyxis* sp. Samples 555-32,CC, 555-41,CC, 555-48,CC, 555-58,CC, 555-59,CC, 555-61,CC, and 555-67,CC contain undiagnostic pyritized diatom frustules. Figure 9 shows that an average sedimentation accumulation rate of 2.5 cm/1000 yr. occurs for the middle and late Miocene of Hole 555.

PALEOCEANOGRAPHY

The eight holes drilled during Leg 81 can be correlated by diatom datums (see Fig. 10). With the exception of Hole 552A, which was hydraulically piston cored, all holes were rotary drilled. Diagnostic diatoms occur in the lower Eocene to Holocene sediments.

With the exception of an unconformity in the Pliocene of Hole 555 (see Backman, this volume), all recognized unconformities are early Miocene or older in age. Unconformities are concentrated in the middle Eocene



Figure 9. Sediment accumulation rate curve calculated for Hole 555 from diatom datum levels.

to middle Miocene sediments of Hole 553A. Samples observed within this interval are partially dissolved of diatoms. However, the presence of *Actinocyclus ingens* and *Rocella gelida* in this interval allows recognition of upper Oligocene to middle Miocene sediments. The concentration of unconformities within this 30-m interval may reflect the subsidence of the Iceland Faroe Ridge and erosion of sediments at Hole 553A by the southward-flowing Norwegian Sea Bottom Water.

Two distinct intervals of dissolution are present within Leg 81 material. The oldest occurs in the late Miocene Nitzschia porteri-Nitzschia miocenica Interval and the youngest occurs in the late Pliocene to Pleistocene Nitzschia marina and Pseudoeunotia doliolus Zones. With the exception of middle Eocene sediments at Holes 552, 552A, and 553A, and middle to upper Miocene sediments at Hole 555, samples examined below the base of the Thalassiosira convexa Zone are generally barren of diatoms. This abrupt change in diatom preservation from samples containing a well-preserved assemblage above to samples dissolved or barren of diatoms below correlates with the interval between the first occurrence of T. praeconvexa and the first occurrence of T. convexa and T. miocenica. These datums allow this preservational change to be correlated with the uppermost portion of Epoch 6 and assigned an age of approximately 6.2 m.y. ago. This assigned age corresponds to the age suggested by Haq et al. (1980) for a late Miocene marine carbonisotope shift. This carbon shift, which is a decrease in the concentration of benthic foraminiferal ¹³C, is recorded in sediments from the Pacific (Keigwin, 1979; Bender and Keigwin, 1979), the Indian (Vincent et al., 1980), and the South Atlantic (Haq et al., 1980) oceans. At present this carbon shift is not recorded in North Atlantic sediment (Shackleton and Cita, 1979; Keigwin, pers. comm., 1984).

The change in diatom preservation within the late Miocene of the Rockall Plateau Region may reflect an increase in the nutrient supply within the surface waters resulting from changes in oceanic circulation. Such changes are conclusive to an increase in phytoplankton productivity, resulting in an increase in the supply rate of diatom frustules to the ocean floor, which would be reflected by an increase in diatom preservation.

The second interval of dissolution occurs within the Pliocene-Pleistocene, *Nitzschia marina*, and *Pseudo-eunotia doliolus* Zones. Samples examined within this interval contain abundant ice-rafted detritus. The first major occurrence of ice-rafted detritus within the lower portion of this interval suggests that the base of this interval approximates the initiation of Northern Hemispheric glaciation within the Rockall Plateau Region. Within the Rockall Plateau Region this event has an age slightly younger than the last occurrence of *N. jouseae* at 2.65 m.y. ago.

Although occasional samples in this dissolution interval contain a well-preserved diatom flora, the majority of samples examined are barren of diatoms. This dissolution interval is included in a detailed study completed on the upper 50 m of sediment recovered from Hole 552A (see Zimmerman et al., this volume).

Figure 11 compares the fluctuation in diatom abundance to the fluctuation in $^{18}O(\%)$, as obtained from the test of benthic foraminifers for the upper 55 m of Hole 552A. The interval from approximately 24 to 29 m (Core 6) is not included within this study because of the disturbed nature of the sediment.

Useful biostratigraphic datums for this study are as follows: Datum 1 (see Fig. 11)-the presence of the silicoflagellate species Mesocena quadrangula and the diatom species N. fossilis in Sample 552A-5-1, 90 cm (19.9 m depth) suggests that this sample is older than the last occurrence of these species, which occurs at approximately 0.79 m.y. ago. This sample occurs at the base of the Jaramillo Event and therefore can be assigned a more precise age of approximately 0.91 m.y. ago. Datum 2 (see Fig. 11)-the last occurrence of T. convexa at Sample 552A-9-1, 63 cm (39.6 m depth) occurs between the Gauss/Matuyama boundary and the Olduvai Event, which indicates an age of approximate 2.2 m.y. ago for this datum. This age agrees with the results of Burckle and Trainer (1979), who assigned an age of 2.2 m.y. ago for this event in Core V28-179 from the central equatorial Pacific. Datum 3 (see Fig. 11)-the last occurrence of N. jouseae in Sample 552A-10-1, 30 cm (40.8 m

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Figure 10. Correlation of age and zonal determinations between Leg 81 holes. (Correlations of zonal boundaries used during Leg 81 are represented by horizontal lines. Solid lines indicate that zonal boundaries are observed in both holes. Dashed lines indicated correlations that are tentative because dissolution and the lack of recovered material prevented accurate placement of zonal boundaries. Shaded areas represent intervals of generally poor preservation.)

depth) falls within the upper portion of the Gauss Paleomagnetic Epoch and has an approximate age of 2.65 m.y. ago. Once again this is in agreement with the results of Burckle and Trainer (1979) (see Fig. 6). Datum A represents the last occurrence of Rhizosolenia curvirostris within the Rockall Plateau Region. Donahue (1970) and Schrader (1973) have estimated the extinction of R. curvirostris to occur in the North Pacific at approximately 0.26 m.y. ago. Kanaya and Koizumi (1970) have assigned an age of 0.25 m.y. ago for this datum. Within the Rockall Region the last occurrence of R. curvirostris in Sample 552A-2-2, 52 cm is older than previously estimated for the North Pacific Region, as it correlates with oxygen isotope Stage 10 in the North Atlantic. Shackleton and Opdyke (1973, 1976) have estimated the isotope 9/10 stage boundary at 0.35 m.y. ago and the 10/11 isotope stage boundary at 0.37 m.y. ago. Therefore, by extrapolation, the last occurrence of R. curvirostris within the Rockall Region occurs at approximately 0.36 m.y. ago. The paleomagnetic results figured here (Fig. 11) for the uppermost portion of Hole 552A are from a study by Kent (see Zimmerman et al., this volume).

The diatom abundance curve (Fig. 11) generally compares with Shackleton's oxygen-isotope curve. Samples with common diatoms compare to samples that have a low ¹⁸O content, whereas samples barren of diatoms correspond to samples with a higher ¹⁸O content. However, this is only true for the interval (Interval 1) above 7.2 m (younger than approximately 0.44 m.y. ago) and the interval (Interval 3) below 33 m (older than approximately 1.8 m.y. ago). The intermediate interval (Interval 2) is generally barren of diatoms.

Within Intervals 1 and 3, samples that are fairly abundant of diatoms correspond to samples representing interglacial conditions. Samples within these intervals that





contain rare (or are barren of) diatoms correspond to samples representing glacial conditions. This can be readily seen by comparing the interglacial oxygen isotope Stages 1, 5, and 11 with samples that have a high diatom abundance. However, isotope Stage 9 does not correspond with the diatom-abundance curve. Likewise, glacial isotope Stages 2 through 10 correlate with samples having rare diatoms or samples that are barren of diatoms.

Similar correspondence occurs in Interval 3. The lowest portion (45–55 m) of this interval is characterized by comparatively small isotope fluctuations with a similar and somewhat less fluctuating diatom-abundance curve. The O^{18} curve first begins to have larger variability at approximately a depth of 44 m (*Datum B*), with the first major isotope shift occurring at a depth of 42 m. This shift may represent the initiation of northern hemispheric glaciation which would have an approximate age of 2.4 m.y. ago within the Rockall Region. At exactly the same depth, a major shift occurs from samples abundant in diatoms (preglacial) to samples barren of diatoms at the glacial event.

The correspondence within Intervals 1 and 3 (Fig. 11), between samples containing abundant diatoms with interglacial conditions and samples barren of diatoms with glacial conditions, suggests that diatom abundance within the Rockall Region is controlled by the latitudinal migration of surficial water masses during climatic cycling. This interpretation is not new and has been cited by numerous workers involving several microfossil groups (McIntyre et al., 1972; CLIMAP, 1976; Ruddiman, 1977; Ruddiman and McIntyre, 1976, 1981; Ruddiman et al., 1977, 1980).

However, the diatom abundance within Interval 2 (Fig. 11) does not correspond to the isotope curve. Although the majority of samples examined from this interval are barren of diatoms, occasional samples do contain a well-preserved diatom assemblage. Interval 2 extends from the middle of the Olduvai Event to the oxygen isotope Stage 11/12 boundary. Therefore an approximate age of 1.8–0.44 m.y. can be extrapolated for this interval.

It appears reasonable to suggest that the pattern observed in Interval 2 is caused by alterations in productivity and/or preservational factors. These alterations are the probable result of a set of complex changing oceanographic and climatic conditions. Any hypothesis proposed at this time to explain this pattern would be pure speculation because of the very limited data available.

However, one should point out that a "glacial sequence" that may be similar to Interval 2 is recorded by Schrader and Fenner (1976) at Sites 336, 346, 348, and 349 in the Norwegian Sea. Samples examined from these sequences are typically barren of diatoms but contain abundant terrigenous material. Occasional samples are recorded that contain a warm temperate diatom assemblage. Schrader and Fenner (1976) suggested these occasional samples represent periods when North Atlantic Drift waters penetrated into the Norwegian Sea Region.

Further studies are required to determine the relationship between these "glacial sequences" from the Norwegian Sea and Interval 2 from the Rockall Region. A positive relationship between these sequences would suggest that these intervals are the result of wide geographic changes rather than the result of local isolated events.

CONCLUSION

A middle Miocene to Holocene diatom zonation is developed from material recovered from the Rockall Plateau Region. This zonation is partially adapted from the diatom zonation defined by Burckle (1972) for the equatorial Pacific and is correlated to both the low-latitude Atlantic and Pacific oceans.

Two intervals of dissolution are recognized within the Rockall Region. A late-Miocene interval is associated with the base of the *Thalassiosira convexa* Zone and is present at all sites. The top of this interval correlates with the uppermost portion of paleomagnetic Epoch 6 and is assigned an age of approximately 6.2 m.y ago. This preservational change may correspond to alteration of oceanic circulation resulting in changes in the rate of productivity.

An interval of dissolution is recognized in Pliocene-Pleistocene material examined from Hole 552A. Diatom abundance in Hole 552A typically corresponds to fluctuating climatic conditions. Diatoms are most abundant during interglacial intervals and rare or absent during glacial intervals. However, diatom abundance in the Rockall Region for the interval from approximately 1.8 to 0.44 m.y. ago does not correspond to climatic fluctuations. Samples examined within this interval are typically barren of diatoms. However, rare samples were observed to contain a well-preserved diatom assemblage.

Although further work is required to explain the Pliocene-Pleistocene Interval, this interval may correspond to a "glacial sequence" recognized by Schrader and Fenner (1976) within the Norwegian Sea.

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APPENDIX List of Flora

- Actinocyclus curvatulus Janisch 1878 in Schmidt et al. (1874-1959) pl. 57, fig. 31; Hustedt 1958, p. 129-130, pl. 8, fig. 81. (Plate 1, Fig.
- 9; Plate 2, Fig. 8). Actinocyclus divisus (Grunow) Hustedt 1958, p. 129, pl. 8, fig. 81;
- synonym: Coscinodiscus divisus Grunow 1884, p. 31, pl. 4, fig. 16. (Plate 2, Fig. 7; Plate 9, Fig. 3).
- Actinocyclus ehrenberg Ralfs in Pritchard 1861, p. 834; Hustedt 1929, p. 525, fig. 298. (Plate 7, Fig. 3).
- Actinocyclus ellipticus Grunow in Van Heurck 1883, pl. 124, fig. 10; Schrader 1973, pl. 8, figs. 7-9, 12-14, 16, 17. (Plate 6, Figs. 4, 7).
- Actinocyclus ellipticus var. elongatus (Grunow) Kolbe 1954, pl. 3, fig. 28.
- Actinocyclus ingens Rattray 1890a s. ampl., p. 149, pl. 11, fig. 7; Baldauf and Barron 1980, pl. 1, figs. 1-4; Baldauf and Barron 1982, p. 68, pl. 1, figs. 6-10. (Plate 7, Figs. 1, 2, 5, and 7).
- **Remarks.** Although a wide morphological variation of this species is observed, for the present paper, all specimens are recognized as *A. ingens* s. ampl. One distinct variation includes specimens that are more robust, characterized by a distinct central hyaline area (see Plate 7, Figs. 5 and 7). The stratigraphic range of this morphological variation is equivalent to the ranges of other morphotypes. However, this variation seems to be restricted geographically to Atlantic Ocean sediments. (Plate 7, Figs. 1, 2, 5, 7).
- Actinoptychus undulatus (Bailey) Ralfs in Pritchard 1861, p. 839, pl. 5, fig. 88; synonym: Actinocyclus undulatus Bailey 1842, pl. 2, fig. 11. (Plate 3, Fig. 10).
- Asterolampra marylandica Ehrenberg 1844a, p. 76, fig. 10; Hustedt 1930, p. 485-587, fig. 271. (Plate 1, Fig. 2).
- Asteromphalus elegans Greville 1859, p. 161, pl. 7, fig. 6. (Plate 1, Fig. 3).
- Asteromphalus hookeri Ehrenberg 1844a, fig. 3; Hustedt 1958, p. 127-128; pl. 8, figs. 88-90. (Plate 1, Figs. 1, 4).
- Bacteriastrum hyalinum Lauder 1864, p. 8, pl. 3, fig. 7; Barron 1980b, pl. 2, fig. 10. (Plate 1, Fig. 5).
- Brightwellia coronata (Brightwell) Ralfs in Pritchard 1861, p. 940; Fenner 1977, p. 512, pl. 2, fig. 5; pl. 3, fig. 1; synonym: Craspedodiscus coronatus Brightwell 1860, p. 95, pl. 5, fig. 6; Brightwellia pulchra Grunow in Van Heurck 1883, pl. 128, fig. 9.

Remarks. This species is very rare within Leg-81 material. Specimens observed are generally poorly preserved but are assigned to this species based on having a spiral arrangement of the areolae

within the "coronal ring" and having the areolae outside the "coronal ring" arranged in closely spaced linear radiating rows. See Fenner 1977 for a complete description. (Plate 13, Fig. 6).

- Cocconeis vitrea Brun 1891, p. 19, pl. 18, fig. 2; Kanaya 1959, p. 110, fig. 6.
- Coscinodiscus lewisianus Greville 1866, p. 78, pl. 8, figs. 8-10; Schrader 1973, p. 703, pl. 8, figs. 1-6, 10, 15. Comments: Very few specimens of this species were observed in Hole 555. (Plate 8, Figs. 7, 8).
- Coscinodiscus marginatus Ehrenberg 1841a, p. 142; Ehrenberg 1854, pl. 18, fig. 44; Hustedt 1930, p. 416, fig. 223. (Plate 8, Figs. 1-3).
- Coscinodiscus nodulifer Schmidt 1878 in Schmidt et al. (1874-1959) Kanaya 1971, pl. 40.3, figs. 1-4. (Plate 3, Fig. 16; Plate 8, Fig. 6).
- Coscinodiscus nodulifer var. cyclopus Jousé 1977, pl. 45, figs. 11, 15; pl. 48, figs. 1-6, pl. 52, figs. 6-7, pl. 77, fig. 17, pl. 78, figs. 1-4, pl. 79, fig. 16; Barron 1980b, pl. 4, fig. 3. (Plate 3, Figs. 17, 18).
- Coscinodiscus oblongus Greville 1866, p. 4, pl. 1, figs. 9, 10; Hustedt 1939, p. 426-427, fig. 229.
- Coscinodiscus obscurus Schmidt 1878, in Schmidt et al. (1874–1959) pl. 61, fig. 16; Rattray 1890b, p. 513, fig. 65; Hustedt 1928, p. 418, pl. 224.
- Coscinodiscus oculus-iridis Ehrenberg 1839, p. 147; Ehrenberg 1854, pl. 18, fig. 42, pl. 19, fig. 2.
- Coscinodiscus plicatus Grunow 1878 in Schmidt et al. (1874–1959), pl. 59, fig. 1; Grunow 1884, p. 73, pl. 3c, fig. 10; Schrader 1973, p. 703, pl. 6, fig. 23.
- Coscinodiscus praenitidus Fenner in Schrader and Fenner 1976, p. 972, pl. 15, fig. 7, pl. 17, fig. 8. (Plate 10, Fig. 4).
- Coscinodiscus praeyabei Schrader 1973, pl. 6, fig. 16, pl. 7, figs. 17-20, 22, 23; Baldauf and Barron, 1982, pl. 3, fig. 4.
- Coscinodiscus radiatus Ehrenberg 1839, p. 148, pl. 3, figs. 1a-c; Hustedt 1930, p. 420, fig. 225.
- Coscinodiscus stellaris Roper 1858, p. 21, pl. 3, fig. 3; Hustedt 1930, p. 396-398, fig. 207; Schrader 1973, pl. 26, fig. 7.
- Coscinodiscus tabularis Grunow 1884, p. 86; Rattray 1890b, p. 583, fig. 135; Baldauf and Barron 1982, p. 69, pl. 4, figs. 6, 7. (Plate 3, Fig. 11).
- Coscinodiscus yabei Kanaya 1959, p. 86, pl. 5, figs. 6-9; Schrader 1973, p. 704, pl. 6, figs. 1-6. (Plate 2, Fig. 9; Plate 8, Fig. 5).
- Coscinodiscus sp. 1, (Plate 8, Fig. 4).
- Craspedodiscus coscinodiscus Ehrenberg 1844b, p. 266; Ehrenberg 1854, pl. 18, fig. 10; Rattray 1890b, p. 600; Kanaya 1971, p. 555, pl. 40.4, figs. 1-3. (Plate 9, Fig. 6).
- Cussia paleacea (Grunow) Schrader 1974b, p. 543, pl. 1, figs. 11-14, synonym: Stoschia paleacea Grunow in Van Heurck 1883, pl. 128, fig. 6.
- Denticulopsis dimorpha (Schrader) Simonsen 1979, p. 64. Synonym: Denticula dimorpha Schrader, 1973, p. 704, pl. 1, figs. 37-46; Barron 1980a, pl. 1, fig. 21.
- Denticulopsis hustedtii (Simonsen and Kanaya) Simonsen 1979, p. 64;
 Baldauf and Barron 1982, pl. 7, figs. 7, 8. synonym: Denticula hustedtii Simonsen and Kanaya 1961, p. 501, pl. 1, figs. 19-25, pl. 2, figs. 36-47; Schrader 1973, p. 704, pl. 2, figs. 28-34, 36-47. (Plate 3, Figs. 3-5).
- Denticulopsis hyalina (Schrader) Simonsen 1979, p. 64; Baldauf and Barron 1982, pl. 7, figs. 2, 4; synonym: Denticula hyalina Schrader 1973, p. 704, pl. 1, figs. 12-22. (Plate 3, Figs. 2, 6-8).
- Denticulopsis nicobarica (Grunow) Simonsen 1979, p. 65; Baldauf and Barron 1982, pl. 7, fig. 6; synonym: Denticula nicobarica Grunow 1868, p. 97, pl. 1a, fig. 5. (Plate 3, Fig. 9).
- Denticulopsis praedimorpha (Akiba, 1979) Barron, 1981, p. 529, pl. 4, figs. 8–10; synonym: Denticula praedimorpha (Akiba, 1979) Barron 1980a, pl. 1, figs. 18–20.
- Denticulopsis punctata (Schrader) Simonsen 1979, p. 65; Baldauf and Barron 1982, pl. 7, fig. 7. synonym: Denticula punctata Schrader 1973, p. 705, pl. 1, figs. 25-30, pl. 3, figs. 16, 17.
- Denticulopsis puncatata var. hustedtii (Schrader) Simonsen 1979; Barron 1981, pl. 4, fig. 1; synonym: Denticula punctata var. hustedtii Schrader 1973, p. 705, pl. 1, figs. 23, 24. (Plate 3, Figs. 1, 15).
- Denticulopsis seminae (Simonsen and Kanaya) Simonsen, 1979; Barron 1981, p. 529. synonym: Denticula seminae Simonsen and Kanaya 1961, p. 503, pl. 1, figs. 26–30; Barron 1980a, pl. 1, fig. 1. (Plate 3, Figs. 12–14).

- Denticulopsis seminae var. fossilis (Koizumi) Simonsen 1979; synonym: Denticula seminae var. fossilis Schrader, Koizumi 1975b, pl. 1, figs. 3, 4.
- Ethmodiscus rex (Wallich) Hendey in Wiseman and Hendey 1953, p. 51, pl. 1, fig. 2; synonym: Coscinodiscus rex Wallich in Rattray 1890b, p. 568, fig. 120.
- Goniothectum odontella Ehrenberg 1844a, p. 82; Gombos and Ciesielski (1983), pl. 22, fig. 9. (Plate 11, Fig. 3).
- Hemidiscus cuneiformis Wallich 1860, p. 42, pl. 2, figs. 3, 4; Barron 1980b, pl. 1, figs. 9, 10. (Plate 1, Figs. 6, 7).
- Hemiaulus polymorphus Grunow 1884, p. 66; Fenner 1977, p. 522, pl. 21, fig. 11, pl. 23, figs. 10, 11, pl. 22, fig. 13.
- Hemiaulus polycistinorum Ehrenberg 1854, pl. 36, figs. 43a, b; Fenner 1977, p. 521, pl. 21, figs. 13, 14; pl. 22, figs. 4, 5, 7-10, pl. 23, figs. 1-4. (Plate 12, Figs. 5, 11).
- Hemiaulus sp. 1. (Plate 13, Fig. 1).
- Hemiaulus sp. 2. (Plate 12, Fig. 10).
- Hyalodiscus dentatus Korotkevicz 1964, p. 104, pl. 1, figs. 1-4.
- Macrora stella (Azpeitia) Hanna 1932, p. 196, pl. 12, fig. 7; synonym: *Pyxidicula stella* Azpeitia 1911, p. 150–152, pl. 1, fig. 1. **Remarks.** This species is not a diatom but is useful as a good guide fossil for middle Miocene sediments (see Schrader 1973, and Hanna
- 1932). (Plate 5, Fig. 9). Mediaria splendida Sheshukova-Poretzkaya 1962, p. 210, pl. 2, fig. 5; Baldauf and Barron 1982, pl. 7, fig. 15.
- Melosira cf. architecturalis Brun 1892 in Schmidt et al. (1874–1959), pl. 177, figs. 45–50; Schrader and Fenner 1976, p. 989, pl. 14, fig. 13; pl. 29, fig. 7, 8,; pl. 35, figs. 1–4.
- **Remarks.** Placement of observed specimens into this taxa is only tentative due to the quality of preservation of the specimens. (Plate 10, Fig. 5).
- Melosira clavigera Grunow in Van Heurck 1882, pl. 91, fig. 1;
- Melosira sulcata (Ehrenberg) Kutzing 1844, p. 55, pl. 2, fig. 7; synonym: Gallionolla sulcata Ehrenberg 1838, p. 170, pl. 21, fig. 5. (Plate 6, Fig. 6).
- Navicula lyra Ehrenberg 1841b, p. 419, pl. 1, fig. 1; Hustedt 1964, p. 453-464, figs. 1516-1523.
- Navicula scutellum (O'Mears) O'Mears 1876, p. 396, pl. 33, fig. 14; synonym: Pinnularia scutellum O'Mears 1869, p. 151, pl. 2, fig. 5. Navicula sp. 1. (Plate 6, Fig. 9).
- Nitzschia cf. cylindrica Burckle 1972, p. 239-240. pl. 2, figs. 1-6; Schrader 1973, p. 707, pl. 5, figs. 27, 32, 33.
 - **Remarks.** Observed specimens of this species have wide morphological variation. Two morphologically distinct end members exist; those that are linear with broadly rounded apices. (Plate 4, Fig. 13) and those which have slightly less rounded apices, and a mid region which is slightly wider than the apices (Plate 4, Fig. 18). Both forms have the same stratigraphic range and until further taxonomic work can be completed, are considered to be variations of the same species. (Plate 4, Figs. 13, 18).
- Nitzschia fossilis (Frenguelli) Kanaya in Kanaya and Koizumi 1970; Schrader 1974a, p. 914, pl. 4, figs. 9-11, 24, 25; Barron 1980a, pl. 2, figs. 3, 4. (Plate 4, Figs. 8, 10, 11, 14, 15, 16?, 17?).
- Nitzschia jouseae Burckle 1972, p. 240. pl. 2, figs. 17-20; Schrader 1973, pl. 4, figs. 20, 21, 22-23 (?). (Plate 6, Figs. 13, 14).
- Nitzschia marina Grunow in Cleve and Grunow 1880, p. 70; Grunow in Van Heurck 1881, pl. 57, figs. 26–27; Schrader 1973, pl. 4, figs. 17–19; Schrader 1974a, pl. 5, figs. 1, 2, 5, 14(?).
 - **Remarks.** N. marina is characterized by having parallel sides and slightly pointed apices (Plate 5, Figs. 1, 2). However, wide morphological variation does occur, with forms seeming to blend into a intermediate Nitzschia fossilis-Nitzschia reinholdii category (see Plate 4, Figs. 1-4). (Plate 4, Figs. 1-4; Plate 5, Figs. 1-3).
- Nitzschia miocenica Burckle 1972, p. 240-241, pl. 2, figs. 10-15; Barron 1980a, p. 672, pl. 2, fig. 8; pl. 3, figs. 3, 4.
- Remarks. The specimen found on Plate 6, Fig. 16 corresponds to Nitzschia sp. aff. N. miocenica Burckle of Barron 1980a, pl. 3, fig. 4. (Plate 6, Figs. 15, 16).
- Nitzschia panduriformis Gregory 1857, p. 529, fig. 57; Fenner 1977, p. 525, pl. 32, figs. 27-29. (Plate 5, Fig. 8).
- Nitzschia porteri Frenguelli 1949, p. 116, pl. 1 figs. 33-34; Schrader 1973, p. 707, pl. 5, figs. 35, 36, 43, 44, 39-41.
- Nitzschia praereinholdii Schrader 1973, p. 708, pl. 5, figs. 20, 23, 26; Barron 1981, pl. 4, fig. 16.

- Nitzschia reinholdii Kanaya and Koizumi 1970; Schrader 1973, p. 708, pl. 4, fig. 12-16, pl. 5, figs. 1-9; Barron 1981, pl. 4, fig. 15. (Plate 4, Figs. 5-7; Plate 5, Fig. 4).
- Nitzschia sp. 1. (Plate 4, Fig. 12).
- Nitzschia sp. 2. (Plate 6, Fig. 12).
- Opephora sp. 1. (Plate 4, Fig. 9).
- Podosira bella Strelnikova 1960, p. 34. (Plte 10, Fig. 6).
- Podosira glacialis (Grunow) Jorgensen 1905, p. 97, pl. 6, fig. 7; synonym: Podosira hormeides var. glacialis Grunow 1884, p. 108, pl. 5e, fig. 32.
- Pseudoeunotia doliolus (Wallich) Grunow in Van Heurck 1881, pl. 35, fig. 22; Schrader 1973, p. 708, pl. 4, figs. 1-8; synonym: Synedra doliolus Wallich 1860, p. 48, pl. 2, fig. 19. (Plate 5, Figs. 5-7).
- Pterotheca aculeifera (Grunow) Van Heurck 1896, p. 430, fig. 151; Fenner 1977, p. 526, pl. 17, figs. 8-21; synonym: Pyxilla aculeifera Grunow in Van Heurck 1882, pl. 83, figs. 13, 14. (Plate 10, Figs. 11, 12).
- Pterotheca danica (Grunow) Forti 1909, p. 13; synonym: Stephanogonia danica Grunow 1866, p. 146. (Plate 12, Figs. 8, 9).
- Pterotheca spada Tempère et Brun, in Brun and Tèmpere 1889, p. 50, pl. 1, fig. 7; Schrader and Fenner 1976, p. 994, pl. 41, figs. 4, 5, 12, 13.
- *Pyrgupyxis prolongata* (Brun) Hendey (1969); Brun 1890-1893, p. 176, pl. 2, fig. 7; Fenner 1977, p. 528, pl. 19, fig. 8.
- Pyxilla renticulata Grove and Sturt 1887, p. 145, pl. 13, fig. 50. (Plate 10, Fig. 9).
- Rhaphoneis amphiceros Ehrenberg 1844a, p. 87; Hustedt 1959, p. 174, figs. 680-681; Schrader 1973, p. 708-709, pl. 25, figs. 2, 3.
- Rhaphoneis surirella (Ehrenberg) Grunow in Cleve and Moller 1878, p. 154-155; Hustedt 1959, p. 173-174, fig. 679; Schrader 1973, p. 709, pl. 25, figs. 4, 6; synonym: Zygoceros surirella Ehrenberg 1839, p. 160, pl. 4, fig. 12.
- Rhizosolenia barboi (Brun) Tempère and Peragallo 1908, p. 26, No. 47; Schrader 1973, p. 709, pl. 24, figs. 4, 7; synonym: *Pyxilla* barboi Brun 1894, p. 87, pl. 5, figs. 16, 17 and 23. (Plate 1, Fig. 12).
- Rhizosolenia bergonii Peragallo 1892, p. 110, pl. 15, fig. 5; Schrader 1973, p. 709, pl. 9, figs. 1–5, 7, 8, 10, 12, 22, 23; pl. 10, figs. 24, 29. (Plate 2, Fig. 5).
- Rhizosolenia curvirostris Jousé 1959, p. 48, pl. 2, fig. 17; Donahue 1970, p. 135–136, fig. 6; Schrader 1973, p. 709, pl. 24, figs. 5, 6, 8, 9.
 - **Remarks.** Specimens observed of this species differ in the amount of curvature and in the robustness of the lateral spine. (compare Plate 2, Fig. 6 with Plate 1, Fig. 10). (Plate 1, Fig. 10; Plate 2, Fig. 6).
- Rhizosolenia hebatata Bailey 1856, p. 5, pl. 1, figs. 18, 19.
- *Rhizosolenia hebatata* forma *hiemalis* Gran 1904, p. 527, pl. 27, fig. 9; Schrader 1973, pl. 9, figs. 11, 13-17, 19-21, 24, 25. (Plate 2, Figs. 1, 2, 4).
- Rhizosolenia interposita Hajós 1976, p. 827, pl. 21, fig. 8.
- Rhizosolenia miocenica Schrader 1973, p. 707, pl. 10, figs. 2-6, 9-11;
 Barron 1980a, pl. 4, fig. 8; Schrader and Fenner 1976, p. 996, pl. 9, figs. 5, 11, 13, 14.
- Rhizosolenia praebarboi Schrader 1973, p. 709, pl. 24, figs. 1-3; Barron 1980a, pl. 2, fig. 18. (Plate 1, Fig. 11).
- Rhizosolenia praebergonii Muchina 1965; Burckle 1972, pl. 1, fig. 1; Koizumi 1968, p. 217, pl. 34, figs. 20, 21; Schrader 1973, pl. 10, fig. 7, pl. 9, fig. 6.
- Remarks. This species is very rare within Leg 81 material. Plate 2, Fig. 3).
- Rhizosolenia styliformis Brightwell 1858, p. 95, pl. 5, figs. 5a, b and d; Hustedt 1930, p. 584-588, figs. 333-335; Schrader 1973, pl. 10, figs 1, 20, 21, 18, 19; pl. 9, fig. 9(?).
- Rocella gelida (Mann) Bukry 1978b, p. 788, pl. 5, figs. 1-13; Gombos and Ciesielski (1983), pl. 6, figs. 1-6, pl. 26, fig. 1. (Plate 9, Figs. 7, 8).
- Roperia tesselata (Roper) Grunow in Van Heurck 1883, pl. 118, figs. 6, 7; Barron 1980b, pl. 3, figs. 8, 10. (Plate 6, Figs. 5, 8).
- Rouxia californica Peragallo in Tempère and Peragallo 1910, p. 245, No. 468-469; Hanna 1930, p. 186-188, pl. 14, figs. 6-7; Schrader 1973, p. 710, pl. 3, figs. 18-20, 26.
- Sceptroneis gemmata Grunow 1886, p. 146. (Plate 10, Fig. 10).
- Stephanopyxis eocenicus Hajós 1976, p. 824, pl. 4, figs. 3, 4.

- Stephanopyxis grunowii Grove and Sturt 1887 in Schmidt et al. (1874– 1959), pl. 130, figs. 1-4; Fenner 1977, p. 532, pl. 12, figs. 10, 13. (Plate 10, Fig. 2).
- Stephanopyxis cf. raeana (Castracane) De Toni 1894, p. 1146; synonym: Systephania raeana Castracane 1886, p. 151, pl. 9, fig. 11. (Plate 12, Fig. 2).
- Stephanopyxis turris (Greville et Arnott) Ralfs 1861, p. 826, pl. 5, fig. 74; Fenner 1977, p. 532, pl. 12, figs. 8, 9; synonym: Cresswellia turris Greville et Arnott 1857, p. 538. (Plate 6, Fig. 10).
- Stepanopyxis turris var. cylindrus Grunow 1884, p. 87, fig. 35. (Plate 10, Figs. 3, 13).
- Stephanopyxis sp. 1. (Plate 10, Fig. 1).
- Stephanopyxis sp. 2. (Plate 11, Fig. 5).
- Stephanopyxis sp. 3. (Plate 17, Fig. 3).
- Stictodiscus angulatus Grunow et Moller 1868 in Kitton 1871, p. 168. (Plate 10, Fig. 8).
- Synedra jouseae Sheshukova-Poretzkaya 1962, p. 208, fig. 4; Schrader 1973, pl. 23, figs. 21-23, 25, 28. (Plate 6, Fig. 1).
- Synedra ulna (Nitzsch) Ehrenberg 1838, p. 211, pl. 17, fig. 1; synonym: Bacillaria ulna Nitzsch 1817, p. 99, pl. 5.
- Thalassionema nitzschiodes (Grunow) Van Heurck 1896, p. 319, fig. 75; Schrader 1973, pl. 23, figs. 2, 6, 8, 9, 10, 26, 29, 34, 12-13. (Plate 6, Figs. 2, 3).
- Thalassionema sp. 1. (Plate 6, Fig. 1).
- Thalassiosira convexa Muchina 1965; Schrader 1974a, pl. 2, figs. 1-5, 10-13; Barron 1980b, pl. 8, fig. 1. (Plate 5, Fig. 10; Plate 7, Figs. 4, 8).
- Thalassiosira convexa var. aspinosa Schrader 1974a, pl. 2, figs. 8, 9, 13–21; synonym: Thalassiosira convexa Koizumi 1975a, pl. 4, figs. 15–18. (Plate 7, Figs. 6, 9).
- Thalassiosira eccentrica Cleve 1904, p. 216; Sheshukova-Poretzkaya 1967, p. 141-142, pl. 14, fig. 4; Schrader 1973, pl. 25, fig. 17; pl. 16, figs. 5, 6. (Plate 9, Figs. 1, 4).
- Thalassiosira jacksonii Koizumi et Barron in Koizumi 1980, p. 396, pl. 1, figs. 11-14; Barron 1980a, pl. 6, figs. 2, 6 1d?; synonym: Thalassiosira sp. b Schrader and Fenner 1976, pl. 17, figs. 5, 10. (Plate 5, Fig. 11).

- Thalassiosira leptopus (Grunow) Hasle and Fryxell 1977, figs. 1-14, 94-96; synonym: Coscinodiscus lineatus Ehrenberg; Reinhold 1937, pl. 11, fig. 7. (Plate 1, Fig. 8; Plate 9, Fig. 5).
- Thalassiosira miocenica Schrader 1974a, p. 916, pl. 22, figs. 1-5, 11-13; Barron 1980a, pl. 3, fig. 6; synonym: Thalassiosira usatchevii Jousé in Burckle 1972, p. 228.
- Thalassiosira oestrupii (Ostenfeld) Proshkina-Lavrenko 1949; Hasle 1960, p. 8, pl. 1, figs. 5, 7, and 1; Schrader 1974a, pl. 1, figs. 3–11, 13–16, 19, 20; pl. 14, fig. 5; synonym: Coscinosira oestrupii Ostenfeld 1900, p. 52. (Plate 2, Figs. 11, 12).
- Thalassiosira praeconvexa Burckle 1972, p. 242-243, pl. 2, figs. 7-9; Schrader 1973, pl. 11, figs. 10-15.
- Thalassiosira sp. 1. (Plate 2, Fig. 10).
- Thalassiosira sp. 2. (Plate 2, Fig. 13).
- Thalassiosira sp. 3. (Plate 9, Fig. 2).
- Thalassiothrix longissima Cleve et Grunow in Cleve and Moller 1878, No. 118, No. 207; Schrader 1973, pl. 23, figs. 7, 17, 18.
- Trinacria pileolus (Ehrenberg) Grunow 1884, p. 68, pl. B, figs. 59, 60;
 Hustedt 1930, p. 885, fig. 529; Fenner 1977, p. 536, pl. 24, figs.
 16-17; synonym: Triceratium pileolus Ehrenberg 1844c, p. 205.
- Trinacria simulacrum Grove and Sturt 1887, p. 144, pl. 13, fig. 46; Fenner 1977, p. 536, pl. 29, fig. 2, pl. 31, fig. 2. (Plate 11, Fig. 10; Plate 13, Fig. 11).
- Trinacria sp. 1. (Plate 10, Fig. 7).
- Trinacria sp. 2. (Plate 12, Fig. 7).
- Xanthiopyxis ovalis Lohman 1938, p. 91, pl. 20, fig. 6; pl. 22, fig. 12.
- Genus and species indet. 1 (Plate 11, Fig. 1).
- Genus and species indet. 2 (Plate 11, Fig. 6).
- Genus and species indet. 3 (Plate 11, Fig. 8).
- Genus and species indet. 4 (Plate 11, Fig. 9).
- Genus and species indet. 5 (Plate 12, Fig. 1).
- Genus and species indet. 6 (Plate 12, Fig. 4).
- Genus and species indet. 7 (Plate 12, Fig. 6).
- Genus and species indet. 8 (Plate 13, Figs. 5, 7).
- Genus and species indet. 9 (Plate 13, Fig. 8).



Plate 1. 1. Asteromphalus hookeri Ehrenberg. Sample 552A-16-1, 130-131 cm, length 48 μm. 2. Asterolampra marylandica Ehrenberg. Sample 555-19-1, 76-77 cm, length 67 μm. 3. Asteromphalus elegans Greville. Sample 552A-9-3, 93-94 cm, length 88 μm. 4. Asteromphalus hookeri Ehrenberg. Sample 552A-15-3, 100-101 cm, length 39 μm. 5. Bacteriastrum hyalinum Lander. Sample 552A-10-2, 130-131 cm, length 10 μm. 6, 7. Hemidiscus cuneiformis Wall; (6) Sample 552A-15-3, 100-101 cm, length 76 μm; (7) Sample 552A-15-3, 100-101 cm, length 56 μm.
8. Thalassiosira leptopus (Grunow) Hasle and Fryxell. Sample 552A-16-1, 130-131 cm, length 53 μm. 9. Actinocyclus curvatulus Janisch. Sample 552A-9-3, 93-94 cm, length 42 μm. 10. Rhizosolenia curvirostris Jousé. Sample 552A-2-2, 90 cm, length 88 μm. 11. Rhizosolenia praebarboi Schrader. Sample 552A-15-3, 100-103 cm, length 67 μm.



Plate 2. 1, 2, 4. Rhixosolenia hebatata forma hiemalis Grun; (1) Sample 552A-15-3, 100-101 cm, length 36 μm; (2) Sample 552A-15-3, 100-101 cm, length 48 μm; (4) Sample 552A-16-1, 130-131, 76 μm. 3. Rhizosolenia praebergonii Schrader. Sample 552A-10-1, 70 cm, length 65 μm. 5. Rhizosolenia bergonii Peragallo. Sample 552A-9-3, 33-34 cm, length 83 μm. 6. Rhizosolenia curvirostris Jousé. Sample 552A-2-1, 90 cm, length 70 μm. 7. Actinocyclus divisus (Grunow) Hustedt. Sample 552A-16-1, 130-131 cm, length 65 μ. 8. Actinocyclus curvatulus Janisch. Sample 552A-15-3, 100-101 cm, length 13 μm. 9. Coscinidiscus yabei Kanaya. Sample 555-19-1, 76-77 cm, length 39 μm. 10. Thalassiosira sp. 1. Sample 552A-16-1, 130-131 cm, length 14 μm. 11, 12. Thalassiosira oestrupii (Ostenfeld) Proskina-Lavrenko; (11) Sample 552A-9-3, 33-34 cm, length 8 μm. 13. Thalassiosira sp. 2. Sample 553A-1, CC, length 8 μm.



Plate 3. 1, 15. Denticulopsis punctata var. hustedtii (Schrader) Simonsen; (1) Sample 555-12-2, 62–63 cm, length 27 μ m; (15) Sample 555-12-2, 62–63 cm, length 39 μ m. 2, 6, 7, 8. Denticulopsis hyalina (Schrader) Simonsen; (2) Sample 552A-29, CC length 37 μ m; (6) Sample 555-19-1, 76–77 cm, length 33 μ m; (7) Sample 555-19-1, 76–77 cm, length 21 μ m; (8) Sample 555-19-1, 76–77 cm, length 14 μ m. 3, 4, 5. Denticulopsis hustedtii (Simonsen and Kanaya) Simonsen; (3) Sample 555-13, CC, length 33 μ m; (4) Sample 555-19-1, 76–77 cm, length 33 μ m. (5) Sample 555-12-2, 62–63 cm, length 28 μ m. 9. Denticulopsis nicobarica (Grunow) Simonsen. Sample 555-21-2, 121–122 cm, length 10 μ m. 10. Actinoptychus undulatus (Bailey) Ralfs. Sample 552A-29, CC, length 17 μ m. 11. Coscinodiscus tabularis Grunow. Sample 555-19-1, 76–77 cm, length 21 μ m; (13) Sample 552A-5-1, 90 cm, length 21 μ m. 12–14. Denticulopsis seminae (Simonsen and Kanaya) Simonsen; (12) Sample 552A-5-1, 90 cm, length 17 μ m; (13) Sample 552A-5-1, 90 cm, length 39 μ m. 16. Coscinodiscus nodulifer Schmidt. Sample 552A-10-2, 110–113 cm, length 39 μ m. 17, 18. Coscinodiscus nodulifer var. cyclopus Jousé; (17) Sample 552A-16-1, 130–131 cm, length 53 μ m; (18) Sample 552A-15-3, 100–101 cm, length 33 μ m.



Plate 4. 1-4. Nitzschia marina-Nitzschia reinholdii; (1) Sample 552A-10-2, 34-35 cm, length 96 μm; (2) Sample 552A-15-3, 100-101 cm, length 100 μm; (3) Sample 552A-10-2, 130-133 cm, length 79 μm; (4) Sample 552A-16-1, 130-131 cm, length 88 μm. 5-7. Nitzschia reinholdii Kanaya and Koizumi; (5) Sample 552A-15-3, 100-101 cm, length 67 μm; (6) Sample 552A-10-2, 130-133 cm, length 56 μm; (7) Sample 552A-10-3, 40-43 cm, length 66 μm. 8, 10, 11, 14, 15. Nitzschia fossilis (Frenguelli) Kanaya; (8) Sample 552A-5-1, 90 cm, length 62 μm; (10) Sample 552A-15-3, 100-101 cm, length 45 μm; (14) Sample 552A-5-1, 90 cm, length 62 μm; (15) Sample 552A-15-3, 100-101 cm, length 45 μm; (14) Sample 552A-5-1, 90 cm, length 21 μm; (15) Sample 552A-15-3, 100-101 cm, length 45 μm; (14) Sample 552A-5-1, 90 cm, length 21 μm; (15) Sample 552A-15-3, 95-96 cm, length 33 μm. 16, 17. Nitzschia cf. fossilis (Frenguelli) Kanaya; (16) Sample 552A-19-3, 95-96 cm, length 33 μm. 9. Opephora sp. 1. Sample 552A-16-1, 130-131 cm, length 106 μm. 12. Nitzschia sp. 1. Sample 552A-9-3, 33-34 cm, length 28 μm. 13, 18. Nitzschia cf. cylindrica Burckle; (13) Sample 552A-19-3, 33-34 cm, length 31 μm; (18) Sample 552A-19-3, 95-96 cm, length 28 μm.



Plate 5. 1, 2. Nitzschia marina Grunow; (1) Sample 552A-15-3, 100-101 cm, length 126 μm; (2) Sample 552A-16-1, 130-131 cm, length 117 μm.
3. Nitzschia cf. marina Grunow. Sample 552A-15-3, 100-101 cm, length 88 μm.
4. Nitzschia reinholdi Kanaya and Koizumi. Sample 552A-2, CC, length 61 μm.
5-7. Pseudoeunotia doliolus (Wallich) Grunow; (5) Sample 552A-2, CC, length 50 μm; (6) Sample 552A-2, CC, length 42 μm; (7) Sample 552A-2, CC, length 25 μm.
8. Nitzschia panduriformis Gregory. Sample 552A-19-3, 95-96 cm, length 47 μm.
9. Macrora stella (Azpeitia) Hanna. Sample 555-19-1, 76-77 cm, length 9 μm.
10. Thalassiosira convexa Muchina. Sample 552A-10-2, 130-133 cm, length 25 μm.
11. Thalassiosira cf. jacksonii Koizumi and Barron. Sample 552A-15-3, 100-101 cm, length 36 μm.
12. Triceratium cinnamomeum Greville. Sample 552A-16-1, 130-131 cm, length 38 μm.



Plate 6. 1. Synedra jouseana Sheshukova-Poretzkaya. Sample 553A-8, CC, length 190 μm. 2, 3. Thalassionema nitzschoides Grunow; (2) Sample 552A-10-1, 130-133 cm, length 84 μm; (3) Sample 552A-15-3, 100-101 cm, length 80 μm. 4, 7. Actinocyclus ellipticus Grunow; (4) Sample 552A-19-3, 95-96 cm, length 28 μm; (7) Sample 552A-25-2, 110-111 cm, length 58 μm. 5, 8. Roperia tesselata (Roper) Grunow; (5) Sample 552A-5-1, 90 cm, length 34 μm; (8) Sample 552A-5-1, 90 cm, length 28 μm. 6. Melosira sulcata (Ehrenbergii) Kutzing. Sample 553A-11-5, 42-43 cm, length 14 μm. 9. Navicula sp. 1. Sample 552A-10-3, 61-62 cm, length 9 μm. 10. Stephanopyxis turris (Greville and Arnott) Ralfs. Sample 552A-10-2, 130-133 cm, length 28 μm. 11. Thalassionema sp. 1. Sample 552A-10-2, 34-35 cm, length 36 μm. 12. Nitzschia sp. 2. Sample 552A-15-3, 100-101 cm, length 44 μm. 15. Nitzschia jouseae Burckle; (13) Sample 552A-10-2, 34-35 cm, length 56 μm; (14) Sample 552A-15-3, 100-101 cm, length 31 μm.



Plate 7. 1, 2, 5, 7. Actinocyclus ingens Rattray; (1) Sample 555-17, CC, 42 μm; (2) Sample 408-29-2, 94-96 cm, 37 μm; (5) Sample 553A-7, CC, length 22 μm; (7) Sample 555, 17, CC length 37 μm. 3. Actinocyclus ehrenbergii Ralfs. Sample 552A-19-3, 95-96 cm, length 25 μm. 4, 8. Thalassiosira convexa Muchina; (4) Sample 552A-10-2, 130-133 cm, length 28 μm; (8) Sample 552A-10-2, 34-35 cm, length 31 μm. 6, 9. Thalassiosira convexa var. aspinosa Schrader. (6) Sample 553A-1, CC, length 18 μm; (9) Sample 552A-10-2, 130-133 cm, length 31 μm.



Plate 8. 1-3. Coscinodiscus marginatus Ehrenberg; (1) Sample 552A-10-2, 110-113 cm, length 109 μm; (2) Sample 552A-9-3, 33-34 cm, length 28 μm; (3) Sample 552A-9-4, 23-24 cm, length 22 μm.
4. Coscinodiscus sp. 1. Sample 552A-9-3, 33-34 cm, length 31 μm.
5. Coscinodiscus yabei Kanaya. Sample 555-19-1, 76-77 cm, length 18 μm.
6. Coscinodiscus nodulifer Schmidt. Sample 553A-1-2, 55-56 cm, length 56 μm.
7. Coscinodiscus cf. lewisianus? Greville. Sample 408-29-2, 94-96 cm, length 42 μm.
8. Coscinodiscus cf. lewisianus? Greville. Sample 555-22-1, 106-107 cm, length 62 μm.



Plate 9. 1, 4. Thalassiosira eccentrica (Ehrenberg) Cleve; (1) Sample 552A-10-2, 34–35 cm, length 36 μm; (4) Sample 552A-19-3, 95–96 cm, length 31 μm. 2. Thalassiosira sp. 3. Sample 553A-11-5, 42–43 cm, length 28 μm. 3. Actinocyclus divisus (Grunow) Hasle and Fryxell. Sample 555-19-1, 76–77 cm, length 56 μm. 5. Thalassiosira leptopus (Grunow) Hasle and Fryxell. Sample 553A-1, CC, length 48 μm. 6. Craspedodiscus coscinodiscus Ehrenberg. Sample 555-19-1, 76–77 cm, length 65 μm. 7, 8. Rocella gelida (Mann) Bukry; (7) Sample 553A-9-4, 67–68 cm, length 50 μm; (8) Sample 553A-9-4, 67–68 cm, length 71 μm.



Plate 10. 1. Stephanopyxis sp. 1. Sample 553A-11-5, 42–43 cm, length 33 μm. 2. Stephanopyxis grunowii Grove et Sturt. Sample 553A-11-5, 42–43 cm, length 30 μm. 3, 13. Stephanopyxis turris var. cylindrus Grunow; (3) Sample 553A-11-5, 42–43 cm, length 42 μm; (13) Sample 553A-11-4, 131 cm, length 24 μm. 4. Coscinodiscus praenitidus Fenner. Sample 553A-8, CC, length 13 μm. 5. Melosira cf. architecturalis Brun. Sample 553A-11-4, 131 cm, length 12 μm. 6. Podosira bella Strenlnikova. Sample 553A-11-4, 131 cm, length 8 μm. 7. Trinacria sp. 1. Sample 553A-9, CC, length 20 μm. 8. Stictodiscus angulatus Grunow et Moller. Sample 553A-11-4, 131 cm, length 75 μm. 9. Pyxilla renticulata Grove et Sturt. Sample 553A-11-4, 131 cm, length 113 μm. 10. Sceptroneis gemmata Grunow. Sample 553A-11-4, 131 cm, length 34 μm. 11, 12. Pterotheca aculeifera (Grunow) Van Heurck; (11) Sample 553A-11-4, 131 cm, length 39 μm; (12) Sample 553A-11-4, 131 cm, length 28 μm.



Plate 11. 1. Genus and sp. indet. Sample 553A-1,CC, length 70 μm. 2. Triceratium sp. 1. Sample 553A-11-4, 131 cm, length 28 μm. 3. Goniothecium odontells Ehrenberg. Sample 553A-11-4, 131 cm, length 33 μm. 4. Trinacria excavata var. tatragona Schmidt. Sample 552A, 9,CC, length 44 μm. 5. Stephanopyxis sp. 2. Sample 553A-8,CC, length 25 μm. 6. Genus and species indet. Sample 553A-11-5, 42-43 cm, length 32 μm. 7. Triceratium sp. 2. Sample 553A-11-4, 131 cm, length 37 μm. 8. Genus and species indet. Sample 553A-11-4, 131 cm, length 18 μm. 9. Genus and species indet. Sample 553A-11-4, length 77 μm.



Plate 12. 1. Genus and species indet. Sample 553A-11-5, 42-43 cm, length 48 μ m. 2. Stephanopyxis cf. raeana (Castrance) De Toni. Sample 553A-11-5, 42-43 cm, length 30 μ m. 3. Stephanopyxis sp. 3. Sample 553A-11-4, 131 cm, length 11 μ m. 4. Genus and species indet. Sample 553A-11-4, 131 cm, length 32 μ m. 5, 11. Hemiaulus polycystinorum Ehrenberg; (5) Sample 553A-11-4, 131 cm, length 30 μ m; (11) Sample 553A-11-4, 131 cm, length 40 μ m. 6. Genus and species indet. Sample 553A-11-4, 131 cm, length 5 μ m. 7. Trinacria sp. 1. Sample 553A-11-5-42-43 cm, length 34 μ m. 8, 9. Pterotheca danica (Grunow) Forti. (8) Sample 553A-11-4, 131 cm, length 50 μ m. (9) Sample 553A-11-4, 131 cm, length 44 μ m. 10. Hemiaulus sp. 2. Sample 553A-11-4-131 cm, length 40 μ m.



Plate 13. 1. Triceratium cf. imperator Trunn et Witt. Sample 553A-11-4, 131 cm, length 61 μm. 2. Triceratium sp. 3. Sample 553A-11-4, 131 cm, length 28 μm. 3. Triceratium cf. pulchrum Hustedt. Sample 553A-11-5, 42-43 cm, length 56 μm. 4. Hemiaulus sp. 1. Sample 553A-11-4, 131 cm, length 30 μm. 5. Genus and species indet. Sample 553A-11-4, 131 cm, length 18 μm. 6. Brightwellia cf. coronata (Brightwell) Ralfs. Sample 552A-9, CC, length 70 μm. 7. Genus and species indet. Sample 553A-11-4, 131 cm, length 14 μm. 8. Genus and species indet. Sample 553A-11-4, 131 cm, length 14 μm. 8. Genus and species indet. Sample 553A-11-4, 131 cm, length 14 μm. 8. Genus and species indet. Sample 553A-11-4, 131 cm, length 14 μm. 8. Genus and species indet. Sample 553A-11-4, 131 cm, length 14 μm. 8. Genus and species indet. Sample 553A-11-4, 131 cm, length 552-10, CC, length 62 μm. 10. Triceratium sp. 3. Sample 553A-11-5, 42-43 cm, length 28 μm. 11. Trinacria simulacrum Grove et Sturt. Sample 553A-11-4, 131 cm, length 56 μm.